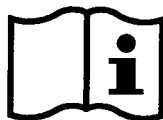


# Product information

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## User's Manual

# Chikungunya Virus IgG capture ELISA

Enzyme immunoassay for the determination of IgG-class antibodies against Chikungunya virus in human serum or plasma



**IB79812**



**96 wells**



**For Research Use Only – Not for Use in Diagnostic Procedures**

## CONTENTS

1. INTENDED USE	3
2. PRINCIPLE OF THE ASSAY	3
3. MATERIALS	3
4. STABILITY AND STORAGE	4
5. REAGENT PREPARATION	4
6. SAMPLE COLLECTION AND PREPARATION	4
7. ASSAY PROCEDURE	5
8. RESULTS	6
9. SPECIFIC PERFORMANCE CHARACTERISTICS	6
10. LIMITATIONS OF THE PROCEDURE	7
11. PRECAUTIONS AND WARNINGS	8
ABBREVIATIONS	8
SUMMARY OF TEST PROCEDURE	9
SYMBOLS USED WITH IBL-AMERICA ASSAYS	12

## 1. INTENDED USE

The Chikungunya Virus IgG capture ELISA is intended for the determination of IgG class antibodies against Chikungunya Virus in human serum or plasma (citrate, heparin). For research use only – Not for use in diagnostic procedures.

## 2. PRINCIPLE OF THE ASSAY

The immunoenzymatic determination of specific IgG-class antibodies is based on the ELISA (Enzyme-linked Immunosorbent Assay) capture technique. Microtiterplates are coated with anti-human IgG antibodies to bind the corresponding antibodies of the sample. After washing the wells to remove all unbound sample material, antigen is added. This antigen binds to the captured specific IgG antibodies. After a further washing step biotinylated antibody is pipetted into the wells. After washing horseradish peroxidase (HRP) labelled streptavidin is added that binds to the captured specific immune complex. After a further washing step the immune complexes are visualized by adding Tetramethylbenzidine (TMB) substrate which gives a blue reaction product. The intensity of this product is proportional to the amount of specific IgG antibodies in the sample. Sulphuric acid is added to stop the reaction. This produces a yellow endpoint colour. Absorbance at 450/620 nm is read using an ELISA Microtiterplate reader.

## 3. MATERIALS

### 3.1. Reagents supplied

- SORB MT Microtiterplate:** 12 break-apart 8-well snap-off strips coated with anti-human IgG; in resealable aluminium foil.
- SAM DIL Sample Dilution Buffer:** 1 bottle containing 100 mL of phosphate buffer (10 mM) for sample dilution; pH 7.2 ± 0.2; coloured yellow; ready to use; white cap; ≤ 0.0015% (v/v) CMIT/ MIT (3:1).
- STOP SOLN Stop Solution:** 1 bottle containing 15 mL sulphuric acid, 0.2 mol/L; ready to use; red cap.
- WASH SOLN 20x Washing Buffer (20x conc.):** 1 bottle containing 50 mL of a 20-fold concentrated phosphate buffer (0.2 M), pH 7.2 ± 0.2, for washing the wells; white cap.
- Ag Antigen, Lyophilized:** 6 bottles containing lyophilized Chikungunya virus antigen solution; red cap; ≤ 0.02% (v/v) MIT.
- Ab SOLN Antibody Solution:** 1 bottle containing 6 mL of biotinylated antibody to Chikungunya virus, ready to use; coloured blue; white cap; ≤ 0.02% (v/v) MIT.
- ENZ CONJ Conjugate:** 1 bottle containing 6 mL Streptavidin conjugated with peroxidase, ready to use; coloured blue; black cap.
- SUB TMB TMB Substrate Solution:** 1 bottle containing 15 mL 3,3',5,5'-tetramethylbenzidine (TMB), < 0.1 %; ready to use; yellow cap.
- CAL C Positive Control:** 1 vial containing 2 mL control; coloured yellow; ready to use; red cap; ≤ 0.02% (v/v) MIT.
- CAL B Cut-off Control:** 1 vial containing 3 mL control; coloured yellow; ready to use; green cap; ≤ 0.02% (v/v) MIT.
- CAL A Negative Control:** 1 vial containing 2 mL control; coloured yellow; ready to use; blue cap; ≤ 0.0015% (v/v) CMIT/ MIT (3:1).

Controls are calibrated in arbitrary units against internal quality control samples, since no international standard reference is available for this assay

\*For hazard and precautionary statements see 12.1

For potential hazardous substances please check the safety data sheet.

### 3.2. Materials supplied

- 1 Cover foil
- 1 Instruction for use (IFU)
- 1 Plate layout

### 3.3. Materials and Equipment needed

- ELISA Microtiterplate reader, equipped for the measurement of absorbance at 450/620 nm
- Incubator 37 °C
- Manual or automatic equipment for rinsing Microtiterplates
- Pipettes to deliver volumes between 10 and 1000 µL
- Vortex tube mixer
- Distilled water
- Disposable tubes

### 4. STABILITY AND STORAGE

Store the kit at 2...8 °C. The opened reagents are stable up to the expiry date stated on the label when stored at 2...8 °C.

### 5. REAGENT PREPARATION

It is very important to bring all reagents and samples to room temperature (20...25 °C) and mix them before starting the test run!

#### 5.1. Microtiterplate

The break-apart snap-off strips are coated anti-human IgG. Immediately after removal of the strips, the remaining strips should be resealed in the aluminium foil along with the desiccant supplied and stored at 2...8 °C.

#### 5.2. Antigen, Lyophilized

The bottles contain lyophilized Chikungunya virus antigen solution. The content of each vial has to be resolved in 1 mL diluted Washing Buffer by turning it slowly (no vortex) and 15 min incubation at room temperature (20...25 °C). The reconstituted solution is stable for 1 day at 2...8 °C.

#### 5.3. Washing Buffer (20x conc.)

Dilute Washing Buffer 1 + 19; e. g. 10 mL Washing Buffer + 190 mL distilled water. The diluted buffer is stable for 5 days at room temperature (20...25 °C). In case crystals appear in the concentrate, warm up the solution to 37 °C e.g. in a water bath. Mix well before dilution.

#### 5.4. TMB Substrate Solution

The reagent is ready to use and has to be stored at 2...8 °C, away from the light. The solution should be colourless or could have a slight blue tinge. If the substrate turns into blue, it may have become contaminated and should be thrown away.

### 6. SAMPLE COLLECTION AND PREPARATION

Use human serum or plasma (citrate, heparin) samples with this assay. If the assay is performed within 5 days after sample collection, the samples should be kept at 2...8 °C; otherwise they should be aliquoted and stored deep-frozen (-70...-20 °C). If samples are stored frozen, mix thawed samples well before testing. Avoid repeated freezing and thawing. Heat inactivation of samples is not recommended.

#### 6.1. Sample Dilution

Before assaying, all samples should be diluted 1+100 with Sample Dilution Buffer. Dispense 10 µL sample and 1 mL Sample Dilution Buffer into tubes to obtain a 1+100 dilution and thoroughly mix with a Vortex.

## 7. ASSAY PROCEDURE

Please read the instruction for use carefully before performing the assay. Result reliability depends on strict adherence to the instruction for use as described. The following test procedure is only validated for manual procedure. If performing the test on ELISA automatic systems we recommend increasing the washing steps from three up to five and the volume of Washing Buffer from 300 µL to 350 µL to avoid washing effects. Pay attention to chapter 12. Prior to commencing the assay, the distribution and identification plan for all samples and standards/controls (duplicates recommended) should be carefully established on the plate layout supplied in the kit. Select the required number of microtiter strips or wells and insert them into the holder.

Perform all assay steps in the order given and without any delays.

A clean, disposable tip should be used for dispensing each standard/control and sample.

Adjust the incubator to  $37 \pm 1$  °C.

1. Dispense 50 µL standards/controls and diluted samples into their respective wells. Leave well A1 for substrate blank.
2. Cover wells with the foil supplied in the kit.
3. **Incubate for 1 hour  $\pm$  5 min at  $37 \pm 1$  °C.**
4. When incubation has been completed, remove the foil, aspirate the content of the wells and wash each well three times with 300 µL of Washing Buffer. Avoid overflows from the reaction wells. The interval between washing and aspiration should be  $> 5$  sec. At the end carefully remove remaining fluid by tapping strips on tissue paper prior to the next step!  
Note: Washing is important! Insufficient washing results in poor precision and false results.
5. Dispense 50 µL reconstituted Chikungunya virus Antigen into all wells except for the Substrate Blank well A1.
6. **Incubate for 30 min at room temperature (20...25 °C).**
7. Repeat step 4.
8. Dispense 50 µL Chikungunya virus Antibody Solution into all wells except for the blank well A1.
9. **Incubate for 30 min at room temperature (20...25 °C).**
10. Repeat step 4.
11. Dispense 50 µL Streptavidin peroxidase conjugate into all wells except for the blank well A1.
12. **Incubate for 30 min at room temperature (20...25 °C).** Do not expose to direct sunlight.
13. Repeat step 4.
14. Dispense 100 µL TMB solution into all wells.
15. **Incubate for exact 15 min. at room temperature (20...25 °C). in the dark.** A blue colour occurs due to an enzymatic reaction.
16. Dispense 100 µL Stop Solution into all wells in the same order and at the same rate as for the TMB Substrate Solution, thereby a colour change from blue to yellow occurs.
17. Measure the absorbance at 450/620nm within 30 min after addition of the Stop Solution.

### 7.1. Measurement

Adjust the ELISA Microtiterplate reader **to zero** using the **Substrate Blank**.

If - due to technical reasons - the ELISA Microtiterplate reader cannot be adjusted to zero using the Substrate Blank, subtract its absorbance value from all other absorbance values measured in order to obtain reliable results! **Measure the absorbance** of all wells at **450 nm** and record the absorbance values for each standard/control and sample in the-plate layout. Bichromatic measurement using a reference wavelength of 620 nm is recommended. **Where applicable calculate the mean absorbance values of all duplicates.**

## 8. RESULTS

### 8.1. Run Validation Criteria

In order for an assay run to be considered valid, these Instructions for Use have to be strictly followed and the following criteria must be met:

- **Substrate Blank:** Absorbance value < **0.100**
- **Negative Control:** Absorbance value < **Cut-off**
- **Cut-off Control:** Absorbance value **0.150 – 1.300**
- **Positive Control:** Absorbance value > **Cut-off**

If these criteria are not met, the test is not valid and must be repeated.

### 8.2. Results

The Cut-off is the mean absorbance value of the Cut-off Control determinations.

Example:

$$\text{Absorbance value Cut-off control } 0.44 + \text{absorbance value Cut-off control } 0.42 = 0.86 / 2 = 0.43$$

$$\text{Cut-off} = 0.43$$

### 9.2.1 Results in Units [U]

$$\frac{\text{Sample (mean) absorbance value} \times 10}{\text{Cut-off}} = [\text{Units} = \text{U}]$$

Example:  $\frac{1.591 \times 10}{0.43} = 37 \text{ U}$

### 8.3. Interpretation of Results

Cut-off	10 U	-
Positive	> 11 U	Antibodies against the pathogen are present. There has been a contact with the antigen (pathogen resp. vaccine).
Equivocal	9 – 11 U	Antibodies against the pathogen could not be detected clearly. It is recommended to repeat the test with a fresh sample in 2 to 4 weeks. If the result is equivocal again the sample is judged as <b>negative</b> .
Negative	< 9 U	The sample contains no antibodies against the pathogen. A previous contact with the antigen (pathogen resp. vaccine) is unlikely.

## 9. SPECIFIC PERFORMANCE CHARACTERISTICS

The results refer to the groups of samples investigated; these are not guaranteed specifications. For further information about the specific performance characteristics please contact IBL-America.

### 9.1. Precision

Intraassay	n	Mean (OD)	CV (%)
#1	24	0.384	7.86
#2	24	1.405	5.15
#3	24	1.877	6.30
Interassay	n	Mean (U)	CV (%)
#1	12	42.70	7.15
#2	12	58.27	8.45
#3	12	2.92	9.56

### 9.2. Specificity

The specificity is defined as the probability of the assay of scoring negative in the absence of the specific analyte. It is 100% (95% confidence interval: 95.89% - 100%).

### 9.3. Sensitivity

The sensitivity is defined as the probability of the assay of scoring positive in the presence of the specific analyte. It is 98.68% (95% confidence interval: 92.89% - 99.97%).

**9.4. Interferences**

Interferences with hemolytic, lipemic or icteric samples are not observed up to a concentration of 10 mg/mL hemoglobin, 5 mg/mL triglycerides and 0.5 mg/mL bilirubin.

**9.5. Cross Reactivity**

Investigation of a sample panel with antibody activities to potentially cross-reacting parameters did not reveal evidence of false-positive results due to cross-reactions. Cross reactivity with antibodies against other alpha viruses cannot be excluded.

**10. LIMITATIONS OF THE PROCEDURE**

Bacterial contamination or repeated freeze-thaw cycles of the sample may affect the absorbance values.

## 11. PRECAUTIONS AND WARNINGS

- The test procedure, the information, the precautions and warnings in the instructions for use have to be strictly followed. The use of the testkits with analyzers and similar equipment has to be validated. Any change in design, composition and test procedure as well as for any use in combination with other products not approved by the manufacturer is not authorized; the user himself is responsible for such changes. The manufacturer is not liable for false results. The manufacturer is not liable for any results by visual analysis of the samples.
- Only for research use.
- All materials of human or animal origin should be regarded and handled as potentially infectious.
- All components of human origin used for the production of these reagents have been tested for anti-HIV antibodies, anti-HCV antibodies and HBsAg and have been found to be non-reactive.
- **The Chikungunya virus antigens are inactivated. All materials should still be regarded and handled as potentially infectious. Wear gloves while performing the test. We recommend using the antigen under BSL2 cabinet (clean bench).**
- Do not interchange reagents or Microtiterplates of different production lots.
- No reagents of other manufacturers should be used along with reagents of this test kit.
- Do not use reagents after expiry date stated on the label.
- Use only clean pipette tips, dispensers, and lab ware.
- Do not interchange screw caps of reagent vials to avoid cross-contamination.
- Close reagent vials tightly immediately after use to avoid evaporation and microbial contamination.
- After first opening and subsequent storage check conjugate and standard/control vials for microbial contamination prior to further use.
- To avoid cross-contamination and falsely elevated results pipette patient samples and dispense reagents without splashing accurately into the wells.
- The ELISA is only designed for qualified personnel following the standards of good laboratory practice (GLP).
- For further internal quality control each laboratory should additionally use known samples.

### 11.1. Safety note for reagents containing hazardous substances

Reagents may contain CMIT/MIT (3:1) or MIT (see 4.1 oder refers to 4.1)  
Therefore, the following hazard and precautionary statements apply.



#### Warning

H317	May cause an allergic skin reaction.
P261	Avoid breathing spray.
P280	Wear protective gloves/ protective clothing.
P302+P352	IF ON SKIN: Wash with plenty of soap and water.
P333+P313	If skin irritation or rash occurs: Get medical advice/ attention.
P362+P364	Take off contaminated and Wash it before reuse.

Further information can be found in the safety data sheet.

### 11.2. Disposal Considerations

Residues of chemicals and preparations are generally considered as hazardous waste. The disposal of this kind of waste is regulated through national and regional laws and regulations. Contact your local authorities or waste management companies which will give advice on how to dispose hazardous waste.

## ABBREVIATIONS

CMIT	5-chlor-2-methyl-4-isothiazolin-3-one
MIT	2-methyl-2H-isothiazol-3-one



**SUMMARY OF TEST PROCEDURE****SCHEME OF THE ASSAY**  
Chikungunya Virus IgG capture ELISA**Test Preparation**

Prepare reagents and samples as described.  
Establish the distribution and identification plan for all samples and standards/controls on the plate layout supplied in the kit.  
Select the required number of microtiter strips or wells and insert them into the holder.












**Assay Procedure**

	Substrate Blank (A1)	Negative control	Cut-off control	Positive control	Sample (diluted 1+100)
Negative control	-	50 µL	-	-	-
Cut-off control	-	-	50 µL	-	-
Positive control	-	-	-	50 µL	-
Sample (diluted 1+100)	-	-	-	-	50 µL
Cover wells with foil supplied in the kit <b>Incubate for 1 h at 37 ± 1 °C</b> Wash each well three times with 300 µL of Washing Buffer					
Reconstituted Antigen	-	50 µL	50 µL	50 µL	50 µL
<b>Incubate for 30 min at room temperature (20...25 °C)</b> Wash each well three times with 300 µL of Washing Buffer					
Antibody Solution	-	50 µL	50 µL	50 µL	50 µL
<b>Incubate for 30 min at room temperature (20...25 °C)</b> Wash each well three times with 300 µL of Washing Buffer					
Streptavidin conjugate	-	50 µL	50 µL	50 µL	50 µL
<b>Incubate for 30 min at room temperature (20...25 °C)</b> Do not expose to direct sunlight Wash each well three times with 300 µL of Washing Buffer					
TMB Substrate Solution	100 µL	100 µL	100 µL	100 µL	100 µL
<b>Incubate for exactly 15 min at room temperature (20...25 °C) in the dark</b>					
Stop Solution	100 µL	100 µL	100 µL	100 µL	100 µL
Photometric measurement at 450 nm (reference wavelength: 620 nm)					





**SYMBOLS USED WITH IBL-AMERICA ASSAYS**

Symbol	English	Deutsch	Français	Espanol	Italiano
	European Conformity	CE-Konformitätskennzeichnung	Conforme aux normes européennes	Conformidad europea	Conformità europea
	Consult instructions for use	Gebrauchsanweisung beachten	Consulter les instructions d'utilisation	Consulte las Instrucciones	Consultare le istruzioni per l'uso
	In vitro diagnostic device	In-vitro-Diagnostikum	Usage Diagnostic in vitro	Diagnóstico in vitro	Per uso Diagnostica in vitro
	For research use only	Nur für Forschungszwecke	Seulement dans le cadre de recherches	Sólo para uso en investigación	Solo a scopo di ricerca
	Catalogue number	Katalog-Nr.	Référence	Número de catálogo	No. di Cat.
	Lot. No. / Batch code	Chargen-Nr.	No. de lot	Número de lote	Lotto no
	Contains sufficient for <n> tests/	Ausreichend für "n" Ansätze	Contenu suffisant pour "n" tests	Contenido suficiente para <n> ensayos	Contenuto sufficiente per "n" saggi
	Note warnings and precautions	Warnhinweise und Vorsichtsmaßnahmen beachten	Avertissements et mesures de précaution font attention	Tiene en cuenta advertencias y precauciones	Annoti avvisi e le precauzioni
	Storage Temperature	Lagerungstemperatur	Temperature de conservation	Temperatura de conservacion	Temperatura di conservazione
	Expiration Date	Mindesthaltbarkeitsdatum	Date limite d'utilisation	Fecha de caducidad	Data di scadenza
	Legal Manufacturer	Hersteller	Fabricant	Fabricante	Fabbricante
<i>Distributed by</i>	Distributor	Vertreiber	Distributeur	Distribuidor	Distributore