## IBL

### **Product information**



Users Manual

# Helicobacter pylori IgG ELISA

Enzyme immunoassay for the detection of human IgG antibodies against Helicobacter pylori in serum and plasma



**IB79236** 



96 wells



For Research Use Only - Not for Use in Diagnostic Procedures

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#### 1. INTENDED USE

The Helicobacter pylori IgG Antibody ELISA Test Kit has been designed for the detection of specific IgG antibodies against Helicobacter pylori in serum and plasma. For research use only, not for use in diagnostic procedures.

#### 2. PRINCIPLE OF THE TESTS

The Helicobacter pylori IgG antibody test kit is based on the principle of the enzyme immunoassay (EIA). Helicobacter antigen is bound on the surface of the microtiter strips. Diluted serum or ready-to-use standards are pipetted into the wells of the microtiter plate. A binding between the IgG antibodies of the serum and the immobilized Helicobacter antigen takes place. After a one hour incubation at room temperature, the plate is rinsed with diluted wash solution, in order to remove unbound material. Then ready-to-use anti-human-IgG peroxidase conjugate is added and incubated for 30 minutes. After a further washing step, the substrate (TMB) solution is pipetted and incubated for 20 minutes, inducing the development of a blue dye in the wells. The color development is terminated by the addition of a stop solution, which changes the color from blue to yellow. The resulting dye is measured spectrophotometrically at the wavelength of 450 nm. The concentration of the IgG antibodies is directly proportional to the intensity of the color.

#### 3. LIMITATIONS, PRECAUTIONS AND GENERAL COMMENTS

- Only for research use! Do not ingest or swallow! The usual laboratory safety precautions as well as
  the prohibition of eating, drinking and smoking in the lab have to be followed.
- All sera and plasma or buffers based upon, have been tested respective to HBsAg, HIV and HCV
  with recognized methods and were found negative. Nevertheless precautions like the use of latex
  gloves have to be taken.
- Serum and reagent spills have to be wiped off with a disinfecting solution (e.g. sodium hypochlorite, 5%) and have to be disposed of properly.
- All reagents have to be brought to room temperature (18 to 25 °C) before performing the test.
- Before pipetting all reagents should be mixed thoroughly by gentle tilting or swinging. Vigorous shaking with formation of foam should be avoided.
- It is important to pipet with constant intervals, so that all the wells of the microtiter plate have the same conditions.
- When removing reagents out of the bottles, care has to be taken that the stoppers are not contaminated. Further a possible mix-up has to be avoided. The content of the bottles is usually sensitive to oxidation, so that they should be opened only for a short time.
- In order to avoid a carry-over or a cross-contamination, separate disposable pipet tips have to be used.
- All reagents have to be used within the expiry period.
- In accordance with a Good Laboratory Practice (GLP) or following ISO9001 all laboratory devices employed should be regularly checked regarding the accuracy and precision. This refers amongst others to microliter pipets and washing or reading (ELISA-Reader) instrumentation.
- The contact of certain reagents, above all the stopping solution and the substrate with skin, eye and mucosa has to be avoided, because possible irritations and acid burns could arise, and there exists a danger of intoxication.

#### 4. REAGENTS PROVIDED

Symbol	Symbol Components	
SORB MT	Helicobacter pylori antigen coated microtiter strips	12
CALA	Calibrator A (Negative Control)	2 mL
CAL B	Calibrator B (Cut-Off Standard)	2 mL
CALC	Calibrator C (Weak Positive Control)	2 mL
CAL D	Calibrator D (Positive Control)	2 mL
ENZ CONJ	Enzyme Conjugate	15 mL
SUB TMB	Substrate	15 mL
STOP SOLN	Stop Solution	15 mL
SAM DIL	Sample Diluent	60 mL
WASH SOLN 10x	Washing Buffer (10×)	60 mL

#### Storage and Stability (refer to the expiry date on the outer box label)

Store kit components at 2-8°C and do not use after the expiry date on the box outer label. Before use, all components should be allowed to warm up to ambient temperature (18-25°C). After use, the plate should be resealed, the bottle caps replaced and tightened and the kit stored at 2-8°C. After the first opening the kit should be used within 3 months, the diluted wash buffer can be kept for 4 weeks at 2-8°C.

#### 4.1. Microtiter Strips

12 strips with 8 breakable wells each, coated with purified natural Helicobacter pylori antigen. Readyto-use.

#### 4.2. Calibrator A (Negative Control)

2 mL, protein solution diluted with PBS, contains no IgG antibodies against Helicobacter. Addition of 0.01% methylisothiazolone and 0.01% bromonitrodioxane. Ready-to-use.

#### 4.3. Calibrator B (Cut-Off Standard)

2 mL human serum diluted with PBS, contains a low concentration of IgG antibodies against Helicobacter. Addition of 0.01 % methylisothiazolone and 0.01 % bromonitrodioxane. Ready-to-use.

#### 4.4. Calibrator C (Weak Positive Control)

2 mL, human serum diluted with PBS, contains a medium concentration of IgG antibodies against Helicobacter. Addition of 0.01 % methylisothiazolone and 0.01 % bromonitrodioxane. Ready-to-use.

#### 4.5. Calibrator D (Positive Control)

2 mL, human serum diluted with PBS, contains a high concentration of IgG antibodies against Helicobacter. Addition of 0.01% methylisothiazolone and 0.01% bromonitrodioxane. Ready-to-use.

#### 4.6. Enzyme Conjugate

15 mL, anti-human-IgG-HRP (rabbit), in protein-containing buffer solution. Addition of 0.01 % methylisothiazolone and 0.01 % bromonitrodioxane and 5 mg/L Proclin™. Ready-to-use.

#### 4.7. Substrate

15 mL, TMB (tetramethylbenzidine). Ready-to-use.

#### 4.8. Stop Solution

15 mL, 1 N acidic solution. Ready-to-use.

#### 4.9. Sample Diluent

60 mL, PBS/BSA buffer. Addition of 0.095 % sodium azide. Ready-to-use.

#### 4.10. Washing Buffer

60 mL, PBS + Tween 20, 10x concentrate. Final concentration: dilute 1+9 with deionized water. If during the cold storage crystals precipitate, the concentrate should be warmed up at 37°C for 15 minutes.

#### 5. MATERIALS REQUIRED BUT NOT PROVIDED

- 5 μL-, 100 μL- and 500 μL micro- and multichannel pipets
- Microtiter Plate Reader (450 nm)
- Microtiter Plate Washer
- Reagent tubes for the serum dilution
- Deionized water
- · Re-usable black lid for covering
- Plastic Bag

#### 6. SAMPLE COLLECTION AND HANDLING

Principally serum or plasma (EDTA, heparin) can be used for the determination. Serum is separated from the blood, which is aseptically drawn by venipuncture, after clotting and centrifugation. The serum or plasma samples can be stored refrigerated (2-8°C) for up to 7 days. For a longer storage they should be kept at -20°C. The samples should not be frozen and thawed repeatedly. Lipemic, hemolytic or bacterially contaminated samples can cause false positive or false negative results.

For the performance of the test the samples (not the standards) have to be diluted 1:101 with ready-to-use sample diluent (e.g.  $5 \mu L$  serum +  $500 \mu L$  sample diluent).

#### 7. ASSAY PROCEDURE

#### 7.1. Preparation of Reagents

**Washing Solution:** dilute before use 1+9 with deionized water. If during the cold storage crystals precipitate, the concentrate should be warmed up at 37°C for 15 minutes.

- Strict adherence to the protocol is advised for reliable performance. Any changes or modifications are the responsibility of the user.
- All reagents and samples must be brought to room temperature before use, but should not be left at this temperature longer than necessary.
- A standard curve should be established with each assay.
- Return the unused microtiter strips to the plastic bag and store them dry at 2-8°C.

#### 7.2. Assay Steps

- 1. Prepare a sufficient amount of microtiter wells for the standards, controls and samples as well as for a substrate blank.
- 2. Pipet 100  $\mu$ L each of the **diluted** (1:101) samples and the **ready-to-use** standards and controls respectively into the wells. Leave one well empty for the substrate blank.
- 3. Cover plate with the re-usable plate cover and incubate at room temperature for 60 minutes.
- 4. Empty the wells of the plate (dump or aspirate) and add 300 μL of diluted washing solution. This procedure is repeated totally three times. Rests of the washing buffer are afterwards removed by gentle tapping of the microtiter plate on a tissue cloth.
- 5. Pipet 100 µL each of ready-to-use conjugate into the wells. Leave one well empty for the substrate blank.
- 6. Cover plate with the re-usable plate cover and incubate at room temperature for 30 minutes.
- 7. Empty the wells of the plate (dump or aspirate) and add 300 µL of diluted washing solution. This procedure is repeated totally three times. Rests of the washing buffer are afterwards removed by gentle tapping of the microtiter plate on a tissue cloth.
- 8. Pipet 100 µL each of the ready-to-use substrate into the wells. This time also the substrate blank is pipetted.
- 9. Cover plate with the re-usable plate cover and incubate at room temperature for 20 minutes in the dark (e.g. drawer).
- 10. To terminate the substrate reaction, pipet 100  $\mu$ L each of the ready-to-use stop solution into the wells. Pipet also the substrate blank.
- 11. After thorough mixing and wiping the bottom of the plate, perform the reading of the absorption at 450 nm (optionally reference wavelength of 620 nm). The color is stable for at least 60 minutes.

#### 8. EVALUATION

#### **Example**

	OD Value	Corrected OD
Substrate Blank	0.015	
Negative Control	0.024	0.009
Cut-Off Standard	0.460	0.445
Weak Positive Control	1.084	1.069
Positive Control	2.213	2.198

#### 8.1. Evaluation (Cut-Off)

The calculated absorptions for the sera, as mentioned above, are compared with the value for the cut-off standard. If the value of the sample is higher, there is a positive result. For a value below the cut-off standard, there is a negative result. It seems reasonable to define a range of +/-20 % around the value of the cut-off as a grey zone. In such a case the repetition of the test with the same serum or with a new sample of the same subject, taken after 2-4 weeks, is recommended. Both samples should be measured in parallel in the same run.

The positive control must show at least the double absorption compared with the cut-off standard.

#### 8.2. Evaluation (U/mL)

The ready-to-use standards and controls of the Helicobacter pylori IgG antibody kit are defined and expressed in arbitrary units (U/mL). Consequently for a given subject follow-up controls become possible. The values for controls and standards in units are printed on the QC data sheet.

For this evaluation the absorptions of the standards and controls are graphically drawn *point-to-point* against their concentrations. From the resulting reference curve the concentration values for each sample can then be extracted in relation to their absorptions. It is also possible to use automatic computer programs. As curve fit *point-to-point* has to be chosen.

Calibrator B with its concentration of 10 U/mL serves as cut-off standard. Analogous to the qualitative evaluation a range of +/-20% around the cut-off is defined as a grey zone. Thus results between 8 and 12 U/mL are reported as borderline.

#### 9. ASSAY CHARACTERISTICS

Helicobacter pylori ELISA	IgG		
Intra-Assay-Precision	8.5 %		
Inter-Assay-Precision	6.3 %		
Inter-Lot-Precision	3.6 – 10.8 %		
Analytical Sensitivity	1.16 U/mL		
Recovery	90 – 93 %		
Linearity	82 – 118 %		
Cross-Reactivity	No cross-reactivity to Yersinia enterocolitis		
Interferences	No interferences to bilirubin up to 0.3 mg/mL,		
	hemoglobin up to 8.0 mg/mL and		
	triglycerides up to 5.0 mg/mL		
Specificity	96 %		
Sensitivity	96 %		

#### 10. REFERENCES

- Cover TL et al. Serologic detection of infection with cagA+ Helicobacter pylori strains. J. Clin. Microbiol., 33: 1496 (1995).
- Cutler AF et al. Accuracy of invasive and non-invasive tests to diagnose Helicobacter pylori infection. Gastroenterology, 109: 136 (1966).
- 3. Dhar R et al. Evaluation and comparison of two immunodiagnostic assays for Helicobacter pylori antibodies with culture results. Diagn. Microbiol. Infect. Dis. 30: 1 (1998).
- 4. Dixon MF. Helicobacter pylori and peptic ulceration; J. Gastroenterol. Hepatol., 6: 125 (1991).
- 5. Donati M et al. Detection of serum antibodies to CagA and VacA and of serum neutralizing activity for vacuolating cytotoxin in patients with Helicobacter pylori-induced gastritis. Clin. Diagn. Lab. Immunol., 4: 478 (1997).
- 6. Evans DJ et al. A sensititve and specific serologic test for detection of Campylobacter pylori infection. Gastroenterology, 96: 1004 (1989).
- Gosciniak G. IgG and IgA antibodies in Helicobacter pylori infections. Zentralbl. Bakteriol., 286:494 (1997).
- 8. Heikkinen M et al. Usefulness of anti-Helicobacter pylori and anti-CagA antibodies in the selection of patients for gastroscopy. Am. J. Gastroenterol., 92: 2225 (1997).
- 9. Karvar S et al. Use of serum-specific immunoglobulins A and G for detection of Helicobacter pylori infection in patients with chronic gastritis by immunoblot analysis. J. Clin. Microbiol., 35: 3058 (1997).

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#### SYMBOLS USED WITH IBL-AMERICA ASSAYS

Symbol	English	Deutsch	Français	Español	Italiano
[]i	Consult instructions for use	Gebrauchsanweisung beachten	Consulter les instruc- tions d'utilisation	Consulte las instruc- ciones de uso	Consultare le istruzioni per l'uso
(€	European Conformity	CE-Konfirmitäts- kennzeichnung	Conformité aux normes européennes	Conformidad europea	Conformità europea
IVD	In vitro diagnostic device	In-vitro-Diagnostikum	Usage Diagnostic in vitro	Para uso Diagnóstico in vitro	Per uso Diagnostica in vitro
RUO	For research use only	Nur für Forschungszwecke	Seulement dans le cadre de recherches	Sólo para uso en investigación	Solo a scopo di ricerca
REF	Catalogue number	Katalog-Nr.	Numéro de catalogue	Número de catálogo	Numero di Catalogo
LOT	Lot. No. / Batch code	Chargen-Nr.	Numéro de lot	Número de lote	Numero di lotto
$\sum$	Contains sufficient for <n> tests/</n>	Ausreichend für "n" Ansätze	Contenu suffisant pour "n" tests	Contenido suficiente para <n> ensayos</n>	Contenuto sufficiente per "n" saggi
1	Storage Temperature	Lagerungstemperatur	Température de conservation	Temperatura de conservación	Temperatura di conservazione
	Expiration Date	Mindesthaltbarkeits- datum	Date limite d'utilisation	Fecha de caducidad	Data di scadenza
<b></b>	Legal Manufacturer	Hersteller	Fabricant	Fabricante	Fabbricante
Distributed by	Distributor	Vertreiber	Distributeur	Distribuidor	Distributore
Content	Content	Inhalt	Conditionnement	Contenido	Contenuto
Volume/No.	Volume / No.	Volumen/Anzahl	Volume/Quantité	Volumen/Número	Volume/Quantità