

## Instruction Manual

### Adenovirus IgA ELISA

**Enzyme immunoassay** for the detection  
of human IgA antibodies against **Adenovirus**  
in serum and plasma

Cat. No.: IB79201  
Storage: 2-8°C

**RUO**

**For Research Use Only – Not for Use in Diagnostic Procedures**

# Product information



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# Product information



## 1. Intended Use

The IBL - AMERICA Adenovirus IgA Antibody ELISA Test Kit has been designed for the detection of specific IgA antibodies against Adenovirus in serum and plasma. For research use only, not for use in diagnostic procedures.

## 2. Discussion

Adenoviruses are ubiquitous pathogens of humans and animals. They are characterized by location inside the cell nucleus, common complement-fixing antigens and marked stability to environmental effects. Adenoviruses are found in all populations throughout the year.. The incubation period is between five and seven days. Adenoviruses mainly infect respiratory and intestinal mucosa, but also the cornea. They are accumulated in the epithelial cells and regional lymph nodes. Adenoviruses cause the widest variety of disorders of the known respiratory viruses.

## 3. Principle of the Test

The IBL - AMERICA Adenovirus IgA antibody test kit is based on the principle of the enzyme immunoassay (EIA). Adenovirus antigen is bound on the surface of the microtiter strips. Diluted sera are pipetted into the wells of the microtiter plate. Binding between the IgA antibodies of the serum and the immobilized Adenovirus antigen takes place. After a one hour incubation at room temperature, the plate is rinsed with diluted wash solution, in order to remove unbound material. Then ready-to-use anti-human-IgA peroxidase conjugate is added and incubated for 30 minutes. After a further washing step, the substrate (TMB) solution is pipetted and incubated for 20 minutes, inducing the development of a blue dye in the wells. The color development is terminated by the addition of a stop solution, which changes the color from blue to yellow. The resulting dye is measured spectrophotometrically at the wavelength of 450 nm.

## 4. Limitations, Precautions and General Comments

- This assay is intended for research use only – not for use in diagnostic procedures.
- Do not ingest or swallow! The usual laboratory safety precautions as well as the prohibition of eating, drinking and smoking in the lab have to be followed.
- All sera and plasma or buffers based upon, have been tested respective to HBsAg, HIV and HCV with recognized methods and were found negative. Nevertheless precautions like the use of latex gloves have to be taken.
- Serum and reagent spills have to be wiped off with a disinfecting solution (e.g. sodium hypochlorite, 5%) and have to be disposed of properly.
- All reagents have to be brought to room temperature (18 to 25 °C) before performing the test.
- Before pipetting all reagents should be mixed thoroughly by gentle tilting or swinging. Vigorous shaking with formation of foam should be avoided.
- It is important to pipet with constant intervals, so that all the wells of the microtiter plate have the same conditions.
- When removing reagents out of the bottles, care has to be taken that the stoppers are not contaminated. Further a possible mix-up has to be avoided. The content of the bottles is usually sensitive to oxidation, so that they should be opened only for a short time.
- In order to avoid a carry-over or a cross-contamination, separate disposable pipet tips have to be used.
- No reagents from different kit lots have to be used, they should not be mixed among one another.
- All reagents have to be used within the expiry period.
- In accordance with Good Laboratory Practice (GLP) or following ISO9001 all laboratory devices employed should be regularly checked regarding the accuracy and precision. This refers amongst others to microliter pipets and washing or reading (ELISA-Reader) instrumentation.

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- The contact of certain reagents, above all the stopping solution and the substrate with skin, eye and mucosa has to be avoided, because possible irritations and acid burns could arise, and there exists a danger of intoxication.
- This assay is manufactured for IBL-America.

## 5. Reagents Provided

Store kit components at 2-8°C and do not use after the expiry date on the box outer label. Before use, all components should be allowed to warm up to ambient temperature (18-25°C). After use, the plate should be resealed, the bottle caps replaced and tightened and the kit stored at 2-8°C. After the first opening the kit should be used within 3 months, the diluted wash buffer can be kept for 4 weeks at 2-8°C.

Components	Volume / Qty.
Adenovirus antigen coated microtiter strips	12 x 8 wells
Calibrator A (Negative Control)	1 x 2 mL
Calibrator B (Cut-Off Standard)	1 x 2 mL
Calibrator C (Weak Positive Control)	1 x 2 mL
Calibrator D (Positive Control)	1 x 2 mL
Enzyme Conjugate	1 x 15 mL
Substrate	1 x 15 mL
Stop Solution	1 x 15 mL
Sample Diluent	1 x 60 mL
Washing Buffer (10×)	1 x 60 mL
Plastic bag	1 ea.

### 5.1. Microtiter Strips

12 strips with 8 breakable wells each, coated with a Adenovirus antigen (Adenovirus Hexon). Ready-to-use.

### 5.2. Calibrator A (Negative Control)

2 mL, protein solution diluted with PBS, contains no IgA antibodies against Adenovirus. Addition of 0.01 % methylisothiazolone and 0.01 % bromonitrodioxane. Ready-to-use.

### 5.3. Calibrator B (Cut-Off Standard)

2 mL human serum diluted with PBS, contains a low concentration of IgA antibodies against Adenovirus. Addition of 0.01 % methylisothiazolone and 0.01 % bromonitrodioxane. Ready-to-use.

### 5.4. Calibrator C (Weak Positive Control)

2 mL, human serum diluted with PBS, contains a medium concentration of IgA antibodies against Adenovirus. Addition of 0.01 % methylisothiazolone and 0.01 % bromonitrodioxane. Ready-to-use.

### 5.5. Calibrator D (Positive Control)

2 mL, human serum diluted with PBS, contains a high concentration of IgA antibodies against Adenovirus. Addition of 0.01 % methylisothiazolone and 0.01 % bromonitrodioxane. Ready-to-use.

### 5.6. Enzyme Conjugate

15 mL, anti-human-IgA-HRP (rabbit), in protein-containing buffer solution. Ready-to-use.

### 5.7. Substrate

15 mL, TMB (tetramethylbenzidine). Ready-to-use.

### 5.8. Stop Solution

15 mL, 0.5 M acidic solution. Ready-to-use.

### 5.9. Sample Diluent

60 mL, PBS/BSA buffer. Addition of 0.095 % sodium azide. Ready-to-use.

### 5.10. Washing Buffer

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60 mL, PBS + Tween 20, 10x concentrate. Final concentration: dilute 1+9 with distilled water. If during the cold storage crystals precipitate, the concentrate should be warmed up at 37°C for 15 minutes.

## 5.11. Plastic Bag

Resealable, for the dry storage of non-used strips.

## 6. Materials Required but not Provided

- 5 µL-, 100 µL- and 500 µL micro- and multichannel pipets
- Microtiter Plate Reader (450 nm)
- Microtiter Plate Washer
- Reagent tubes for the serum dilution
- Bidistilled water
- Re-usable black lid for covering  
(Available upon request at IBL-America)

## 7. Collection and Handling of Unknowns

Principally serum or plasma (EDTA, heparin) can be used in this test. Serum is separated from the blood, which is aseptically drawn by venipuncture, after clotting and centrifugation. Serum or plasma can be stored refrigerated (2-8°C) for up to 48 hours, for longer storage they should be kept at -20 °C. They should not be frozen and thawed repeatedly. Lipemia, hemolysis or bacterial contamination can cause inaccurate results.

All unknowns must be diluted 1:101 with ready-to-use diluent (e.g. 5 µL serum + 500 µL diluent). No other component of the kit requires similar dilution.

## 8. Assay Procedure

### 8.1. Preparation of Reagents

**Wash Solution:** dilute before use 1+9 with distilled water. If during the cold storage crystals precipitate, the concentrate should be warmed up at 37°C for 15 minutes.

- Strict adherence to the protocol is advised for reliable performance. Any changes or modifications are the responsibility of the user.
- All reagents and unknowns must be brought to room temperature before use, but should not be left at this temperature longer than necessary.
- All test materials should be assayed in duplicate.
- Return the unused microtiter strips to the plastic bag and store them dry at 2-8°C.

### 8.2. Assay Steps

1. Prepare a sufficient amount of microtiter wells for all materials in duplicate and also include a substrate blank.
2. Pipet 100 µL each of the **diluted** (1:101) unknowns and the **ready-to-use** calibrators and controls respectively into the wells. Leave the substrate blank empty.
3. Cover plate with the re-usable plate cover and incubate at room temperature for 60 minutes.
4. Empty the wells of the plate (dump or aspirate) and add 300 µL of diluted washing solution. This procedure is repeated totally three times. Remove residual wash buffer by inverting the microtiter plate and tapping briskly on absorbent material.
5. Immediately pipet 100 µL of ready-to-use conjugate into the microwells. Leave the substrate blank microwell empty.
6. Cover plate with the re-usable plate cover and incubate at room temperature for 30 minutes.

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7. Empty the wells of the plate (dump or aspirate) and add 300  $\mu\text{L}$  of diluted washing solution. This procedure is repeated for a total of three times. Remove residual wash buffer by inverting the microtiter plate and tapping briskly on absorbent material.
8. Pipet 100  $\mu\text{L}$  of the ready-to-use substrate into every microwell, including the substrate blank.
9. Cover plate with the re-usable plate cover and incubate at room temperature for 20 minutes.
10. To terminate the substrate reaction, pipet 100  $\mu\text{L}$  of the ready-to-use stop solution into every microwell, including the substrate blank.
11. After thorough mixing and wiping the bottom of the plate, perform the reading of the absorption at 450 nm (optionally reference wavelength of 620 nm). The color is stable for at least 60 minutes.

## 9. Results

The ODs of the calibrators (y-axis, linear) are plotted against their concentration (x-axis, logarithmic) either on semi-logarithmic graph paper or using an automated method. A good fit is provided with cubic spline, 4 parameter logistics or Logit-Log.

The initial dilution of unknowns has been taken into consideration when reading results from the graph. Results of unknowns of higher predilution have to be adjusted for the dilution factor.

Unknowns showing concentrations above the highest calibrator have to be diluted as described in "Assay Procedure" (chapter 8.) and reassayed.

Values suggested by literature:

U/mL	
< 8	negative
8 - 12	equivocal
> 12	positive

In an in-house study, apparently healthy research subjects showed the following results:

Ig Isotype	n	Interpretation		
		positive	equivocal	negative
IgA	51	8.0 %	1.1 %	90.9 %