Product information



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Users Manual



Testosterone ELISA

Enzyme Immunoassay for the quantitative measurement of Testosterone in serum and plasma





IB79106



96 wells

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1 INTRODUCTION

1.1 Intended Use

The **IBL-AMERICA Testosterone ELISA** is an enzyme immunoassay for the quantitative *in vitro diagnostic* measurement of Testosterone in serum and plasma (EDTA, lithium heparin or citrate plasma). **This assay is intended for in vitro diagnostic use only.**

1.2 Summary and Explanation

Testosterone is the major androgenic steroid hormone. It is responsible for the development of the male external genitalia and secondary sexual characteristics. In females, its main role is as an estrogen precursor. In both genders, it also exerts anabolic effects and influences behavior. In men, testosterone is secreted by the testicular Leydig cells and, to a minor extent, by the adrenal cortex. At puberty, testosterone increases dramatically in boys and progressively declines starting between the fourth and sixth decades of life. In premenopausal women, the ovaries are the main source of testosterone with minor contributions by the adrenals and peripheral tissues. After menopause, ovarian testosterone production is significantly diminished. Testosterone production in testes and ovaries is regulated via pituitary-gonadal feedback involving luteinizing hormone (LH) and, to a lesser degree, inhibins and activins. 40-50 % of the total testosterone in blood is bound with high affinity to sex hormone-binding globulin (SHBG), while 40-50 % is bound with low affinity to albumin, and only 1 2 % can be detected as unbound or free testosterone. Free Testosterone and albuminbound Testosterone are considered bioactive.

In men, mild-to-moderate testosterone elevations are usually asymptomatic while high levels of testosterone are associated with hypothalamic-pituitary-unit dysfunction, testicular tumors, congenital adrenal hyperplasia, prostate cancer or intake of anabolic steroids.

Male testosterone levels below the reference range indicate partial or complete hypogonadism, caused either by primary or secondary/tertiary (pituitary/hypothalamic) testicular failure. Low levels of testosterone are encountered in male patients with the following diseases: primary hypogonadism (e.g. Klinefelter's syndrome), testicular feminization, orchidectomy, congenital cryptorchidism, enzymatic defects, anorexia, liver cirrhosis, drug abuse, or prepubescent intake of anabolic steroids. In adult women, excess testosterone production results in varying degrees of virilization, including hirsutism, acne, oligo-amenorrhea. Increased testosterone levels may be caused by polycystic ovaries, ovarian and adrenal tumors, adrenal hyperplasia, Cushing's syndrome, or congenital adrenal hyperplasia. Decreased testosterone in females can be caused by primary or secondary insufficiency of the ovaries, intake of ovulation blockers, liver cirrhosis, drug abuse, Addison's disease or anorexia. Symptoms of low testosterone may include decline in libido and nonspecific mood changes.

2 PRINCIPLE OF THE TEST

The IBL-AMERICA Testosterone ELISA Kit is a solid phase enzyme-linked immunosorbent assay (ELISA) based on the principle of competitive binding.

The microtiter wells are coated with a monoclonal (mouse) antibody directed towards a unique antigenic site of the testosterone molecule. During the first incubation, the testosterone in the added sample competes with the added enzyme conjugate, which is testosterone conjugated to horseradish peroxidase, for binding to the coated antibody. After a washing step to remove all unbound substances, the solid phase is incubated with the substrate solution. The colorimetric reaction is stopped by addition of stop solution, and optical density (OD) of the resulting yellow product is measured. The intensity of color is inversely proportional to the concentration of the analyte in the sample.

A standard curve is constructed by plotting OD values against concentrations of standards, and concentrations of unknown samples are determined using this standard curve.

3 WARNING AND PRECAUTIONS

- This kit is for in vitro diagnostic use only. For professional use only.
- All reagents of this test kit which contain human serum or plasma have been tested and confirmed negative for HIV I/II, HBsAg and HCV by FDA approved procedures. All reagents, however, should be treated as potential biohazards in use and for disposal.
- Before starting the assay, read the instructions completely and carefully. <u>Use the valid version of the package insert provided with the kit.</u> Be sure that everything is understood.
- The microplate contains snap-off strips. Unused wells must be stored at 2 °C to 8 °C in the sealed foil pouch and used in the frame provided.
- Pipetting of samples and reagents must be done as quickly as possible and in the same sequence for each step.
- Use reservoirs only for single reagents. This especially applies to the substrate reservoirs. Using a
 reservoir for dispensing a substrate solution that had previously been used for the conjugate solution may
 turn solution coloured. Do not pour reagents back into vials as reagent contamination may occur.
- Mix the contents of the microplate wells thoroughly to ensure good test results. Do not reuse microwells.
- Do not let wells dry during assay; add reagents immediately after completing the rinsing steps.
- Allow the reagents to reach room temperature (21-26°C) before starting the test. Temperature will affect the absorbance readings of the assay. However, values for the patient samples will not be affected.
- Never pipet by mouth and avoid contact of reagents and specimens with skin and mucous membranes.
- Do not smoke, eat, drink or apply cosmetics in areas where specimens or kit reagents are handled.
- Wear disposable latex gloves when handling specimens and reagents. Microbial contamination of reagents or specimens may give false results.
- Handling should be in accordance with the procedures defined by an appropriate national biohazard safety guideline or regulation.
- Do not use reagents beyond expiry date as shown on the kit labels.
- All indicated volumes have to be performed according to the protocol. Optimal test results are only
 obtained when using calibrated pipette and microtiter plate readers.
- Do not mix or use components from kits with different lot numbers. It is advised not to exchange wells of different plates even of the same lot. The kits may have been shipped or stored under different conditions and the binding characteristics of the plates may result slightly different.
- Avoid contact with Stop Solution containing 1 N acidic solution. It may cause skin irritation and burns.
- Some reagents contain Proclin 300, BND and/or MIT as preservatives. In case of contact with eyes or skin, flush immediately with water.
- TMB substrate has an irritant effect on skin and mucosa. In case of possible contact, wash eyes with an abundant volume of water and skin with soap and abundant water. Wash contaminated objects before reusing them. If inhaled, take the person to open air.
- Chemicals and prepared or used reagents have to be treated as hazardous waste according the national biohazard safety guideline or regulation.
- For information on hazardous substances included in the kit please refer to the Material Safety Data Sheets. Material Safety Data Sheets for this product are available upon request directly from IBL-America.

4 KIT COMPONENTS

4.1 Contents of the Kit

1. **SORB MT Microtiterwells**, 12x8 (break apart) strips, 96 wells;

Wells coated with anti-Testosterone antibody (monoclonal).

2. CAL 0 - 6 Standard (Standard 0-6), 7 vials, 1 ml, ready to use

Concentrations: 0 - 0.2 - 0.5 - 1 - 2 - 6 - 16 ng/ml

Conversion: 1 ng/ml = 3.467 nmol/l;

Contain non-mercury preservative.

3. **ENZ CONJ Enzyme Conjugate**, 1 vial, 25 ml, ready to use;

Testosterone conjugated to horseradish peroxidase

Contains non-mercury preservative.

- 4. SUB TMB Substrate Solution, 1 vial, 25 ml, ready to use; Tetramethylbenzidine (TMB)
- 5. **STOP SOLN Stop Solution**, 1 vial, 14 ml, ready to use; contains 0.5 M H₂SO₄.

Avoid contact with the stop solution. It may cause skin irritations and burns.

6. WASH SOLN 40x Wash Solution, 1 vial, 30 ml (40X concentrated); see "Reagent Preparation".

Note: Additional Standard 0 for sample dilution is available on request.

4.2 Materials required but not provided

- A microtiterplate calibrated reader (450 nm, with reference wavelength at 620 nm to 630 nm)
- Calibrated variable precision micropipettes
- Absorbent paper
- Distilled water
- Timer
- Graph paper or software for data reduction

4.3 Storage Conditions

When stored at 2-8°C unopened reagents will retain reactivity until expiration date. Do not use reagents beyond this date.

Opened reagents must be stored at 2-8°C. Microtiter wells must be stored at 2-8°C. Once the foilbag has been opened, care should be taken to close it tightly again.

Opened kits retain activity for 8 weeks if stored as described above.

4.4 Preparation of Reagents

Allow all reagents and required number of strips to reach room temperature prior to use.

Wash Solution

Add deionized water to the 40X concentrated Wash Solution. Dilute 30mL of concentrated Wash Solution with 1170mL deionized water to a final volume 1200mL.

The diluted Wash Solution is stable for 2 weeks at room temperature.

4.5 Disposal of the Kit

The disposal of the kit must be made according to the national regulations. Special information for this product is given in the Material Safety Data Sheets.

4.6 Damaged Test Kits

In case of any severe damage of the test kit or components, IBL-AMERICA have to be informed written, latest one week after receiving the kit. Severely damaged single components should not be used for a test run. They have to be stored until a final solution has been found. After this, they should be disposed according to the official regulations.

5 SPECIMEN COLLECTION AND PREPARATION

Serum or plasma (EDTA, lithium heparin or citrate plasma) can be used in this assay. Note: Samples containing sodium azide should not be used in the assay. In general, it should be avoided to use haemolytic, icteric, or lipaemic specimens. For further information refer to chapter "Interfering Substances".

5.1 Specimen Collection

Serum:

Collect blood by venipuncture (e.g Sarstedt Monovette # 02.1388.001), allow to clot, and separate serum by centrifugation at room temperature. Do not centrifuge before complete clotting has occurred. Patients receiving anticoagulant therapy may require increased clotting time.

Plasma:

Whole blood should be collected into centrifuge tubes containing anticoagulant (e.g. Sarstedt Monovette with the appropriate plasma preparation) and centrifuged immediately after collection.

5.2 Specimen Storage and Preparation

Specimens should be capped and may be stored for up to 7 days at 2-8°C prior to assaying. Specimens stored for a longer time (up to 12 months) should be frozen only once at -20°C prior to assay. Thawed samples should be inverted several times prior to testing.

5.3 Specimen Dilution

If in an initial assay, a serum specimen is found to contain more than the highest standard, the specimens can be diluted with *Standard* 0 and re-assayed as described in the Assay Procedure. For the calculation of the concentrations this dilution factor has to be taken into account. Example:

a) dilution 1:10: 10 ul Serum + 90 ul Standard 0 (mix thoroughly)

b) dilution 1:100: 10 µl dilution a) 1:10 + 90 µl Standard 0 (mix thoroughly).

6 TEST PROCEDURE

6.1 General Remarks

- All reagents and specimens must be allowed to come to room temperature before use. All reagents must be mixed without foaming.
- Once the test has been started, all steps should be completed without interruption.
- Use new disposal plastic pipette tips for each standard, control, or sample in order to avoid cross contamination.
- Optical density is a function of the incubation time and temperature. Before starting the assay, it is recommended that all reagents are ready, caps removed, all needed wells secured in holder, etc. This will ensure equal elapsed time for each pipetting step without interruption.
- As a general rule, the enzymatic reaction is linearly proportional to time and temperature.

6.2 Test Procedure

Each run must include a standard curve.

- 1. Secure the desired number of Microtiter wells in the frame holder.
- Dispense 25 μI of each Standard, Control and samples with new disposable tips into appropriate
 wells.
- 3. Dispense 200 µl Enzyme Conjugate into each well.
 - Thoroughly mix for 10 seconds. It is important to have a complete mixing in this step.
- 4. Incubate for **60 minutes** at room temperature.
- 5. Rinse the wells **3 times** with **400 μl** diluted *Wash Solution* per well, if a plate washer is used

Briskly shake out the contents of the wells. Rinse the wells **3 times** with **300 µl** diluted *Wash Solution* per well for manual washing. Strike the wells sharply on absorbent paper to remove residual droplets. **Important note:** The sensitivity and precision of this assay is markedly influenced by the correct performance of the washing procedure!

- 6. Add 200 μI of Substrate Solution to each well.
- 7. Incubate for **15 minutes** at room temperature.
- 8. Stop the enzymatic reaction by adding **100 µl** of **Stop Solution** to each well.
- Determine the optical density of the solution in each well at 450 (reading) and at 620 nm to 630 nm (background subtraction, recommended) with a microtiter plate reader.
 It is recommended that the wells be read within 10 minutes after adding the Stop Solution.

6.3 Calulation of Results

- 1. Calculate the average optical density (OD) values for each set of standards, controls and patient samples.
- 2. Using graph paper, construct a standard curve by plotting the mean OD obtained from each standard against its concentration with OD value on the vertical (Y) axis and concentration on the horizontal (X) axis.
- 3. Using the mean OD value for each sample determine the corresponding concentration from the standard curve.
- 4. Automated method: The results in the Instructions for Use have been calculated automatically using a 4-Parameter curve fit. (4-Parameter Rodbard or 4-Parameter Marquardt are the preferred methods.) Other data reduction functions may give slightly different results.
- 5. The concentration of the samples can be read directly from this standard curve. Samples with concentrations higher than that of the highest standard have to be further diluted or reported as > 16.0 ng/mL. For the calculation of the concentrations this dilution factor has to be taken into account.

6.3.1 Example of Typical Standard Curve

The following data is for demonstration only and **cannot** be used in place of data generations at the time of assay.

Standard	Optical Units (450 nm)		
Standard 0 (0 ng/ml)	2.1		
Standard 1 (0.2 ng/ml)	1.71		
Standard 2 (0.5 ng/ml)	1.44		
Standard 3 (1 ng/ml)	1.18		
Standard 4 (2 ng/ml)	0.89		
Standard 5 (6 ng/ml)	0.46		
Standard 6 (16 ng/ml)	0.24		

7 EXPECTED NORMAL VALUES

It is strongly recommended that each laboratory should determine its own normal and abnormal values. In a study conducted with apparently normal healthy adults, using the IBL-AMERICA Testosterone ELISA the following values are observed:

Population	5% Percentile	95% Percentile	
Males	2.0 ng/mL	6.9 ng/mL	
Females	0.26 ng/mL	1.22 ng/mL	

The results alone should not be the only reason for any therapeutic consequences. The results should be correlated to other clinical observations and diagnostic tests.

8 QUALITY CONTROL

Good laboratory practice requires that controls be run with each calibration curve. A statistically significant number of controls should be assayed to establish mean values and acceptable ranges to assure proper performance. It is recommended to use control samples according to state and federal regulations. The use of control samples is advised to assure the day to day validity of results. Use controls at both normal and pathological levels. The controls and the corresponding results of the QC-Laboratory are stated in the QC certificate added to the kit. The values and ranges stated on the QC sheet always refer to the current kit lot and should be used for direct comparison of the results.

It is also recommended to make use of national or international Quality Assessment programs in order to ensure the accuracy of the results.

Employ appropriate statistical methods for analysing control values and trends. If the results of the assay do not fit to the established acceptable ranges of control materials patient results should be considered invalid. In this case, please check the following technical areas: Pipetting and timing devices; photometer, expiration dates of reagents, storage and incubation conditions, aspiration and washing methods.

After checking the above mentioned items without finding any error contact your distributor or IBL-AMERICA directly.

9 PERFORMANCE CHARACTERISTICS

9.1 Assay Dynamic Range

The range of the assay is between 0.083 – 16 ng/ml.

9.2 Specificity of Antibodies (Cross Reactivity)

The following substances were tested for cross-reactivity of the assay:

Analyte	Cross Reactivity
Testosterone	100.0
DHT	12.9
5α-Dihydrotestosterone	0.8
Androstenedione	0.9
11β-Hydroxyestosterone	3.3
17α-Methyltestosterone	0.1
19-Nortestosterone	3.3
DHEA	0.3
DHEA-S	< 0.1
Epitestosterone	< 0.1
Progesterone	< 0.1
Cortisol	< 0.1
Estrone	< 0.1
Estradiol	< 0.1
Estriol	< 0.1
Danazol	< 0.1

9.3 Analytical Sensitivity

The <u>analytical sensitivity</u> of the IBL-AMERICA ELISA was calculated by subtracting 2 standard deviations from the mean of 20 replicate analyses of the Zero Standard (S0) and was found to be 0.083ng/mL.

9.4 Reproducibility

9.4.1 Intra Assay Variation

The within assay variability is shown below:

Sample	n	Mean (ng/ml)	CV (%)
1	20	0.7	4.2
2	20	4.9	3.3
3	20	11.3	3.3

9.4.2 Inter Assay Variation

The between assay variability is shown below:

Sample	n	Mean (ng/ml)	CV (%)
1	20	0.8	9.9
2	20	5.2	6.7
3	20	11.4	4.7

9.5 Recovery

Samples have been spiked by adding testosterone solutions with known concentrations.

The recovery (%) was calculated by multiplying the ratio of measured and expected values with 100.

		Sample 1	Sample 2	Sample 3
Concentration (ng/mL)		1.1	6.1	11.6
Average Recovery (%)		109.2	100.5	109.1
Panga of Pagayany (9/)	from	86.9	92.2	108.1
Range of Recovery (%)	to	110.7	110.1	110.1

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9.6 Linearity

Samples were measured undiluted and in serial dilutions with standard 0. The recovery (%) was calculated by multiplying the ratio of expected and measured values with 100

		Sample 1	Sample 2	Sample 3
Concentration (ng/mL)		1.1	6.1	11.3
Average Recovery (%)		95.5	101.5	104.9
Denge of Deceyany (9/)	from	86.1	89.0	97.9
Range of Recovery (%)	to	106.6	110.6	110.0

10 LIMITATIONS OF USE

Reliable and reproducible results will be obtained when the assay procedure is performed with a complete understanding of the package insert instruction and with adherence to good laboratory practice. Any improper handling of samples or modification of this test might influence the results.

10.1 Interfering Substances

Haemoglobin (up to 4 mg/ml), Bilirubin (up to 0.25 mg/ml) and Triglyceride (up to 7.5 mg/ml) have no influence on the assay results.

10.2 Drug Interferences

Until today no substances (drugs) are known to us, which have an influence to the measurement of Testosterone in a sample.

10.3 High-Dose-Hook Effect

No hook effect was observed in this test.

11 LEGAL ASPECTS

11.1 Reliability of Results

The test must be performed exactly as per the manufacturer's instructions for use. Moreover the user must strictly adhere to the rules of GLP (Good Laboratory Practice) or other applicable national standards and/or laws. This is especially relevant for the use of control reagents. It is important to always include, within the test procedure, a sufficient number of controls for validating the accuracy and precision of the test. The test results are valid only if all controls are within the specified ranges and if all other test parameters are also within the given assay specifications. In case of any doubt or concern please contact IBL-AMERICA.

11.2 Therapeutic Consequences

Therapeutic consequences should never be based on laboratory results alone even if all test results are in agreement with the items as stated under point 11.1. Any laboratory result is only a part of the total clinical picture of a patient.

Only in cases where the laboratory results are in acceptable agreement with the overall clinical picture of the patient should therapeutic consequences be derived.

The test result itself should never be the sole determinant for deriving any therapeutic consequences.

11.3 Liability

Any modification of the test kit and/or exchange or mixture of any components of different lots from one test kit to another could negatively affect the intended results and validity of the overall test. Such modification and/or exchanges invalidate any claim for replacement.

Claims submitted due to customer misinterpretation of laboratory results subject to point 10.2. are also invalid. Regardless, in the event of any claim, the manufacturer's liability is not to exceed the value of the test kit. Any damage caused to the test kit during transportation is not subject to the liability of the manufacturer.

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13 SYMBOLS USED WITH IBL-AMERICA ASSAYS

Symbol	English	Deutsch	Français	Español	Italiano
Ţ <u>i</u>	Consult instructions for use	Gebrauchsanweisung beachten	Consulter les instructions d'utilisation	Consulte las instrucciones de uso	Consultare le istruzioni per l'uso
(€	European Conformity	CE-Konfirmitäts- kennzeichnung	Conformité aux normes européennes	Conformidad europea	Conformità europea
IVD	In vitro diagnostic device	In-vitro-Diagnostikum	Usage Diagnostic in vitro	Para uso Diagnóstico in vitro	Per uso Diagnostica in vitro
RUO	For research use only	Nur für Forschungszwecke	Seulement dans le cadre de recherches	Sólo para uso en investigación	Solo a scopo di ricerca
REF	Catalogue number	Katalog-Nr.	Numéro de catalogue	Número de catálogo	Numero di Catalogo
LOT	Lot. No. / Batch code	Chargen-Nr.	Numéro de lot	Número de lote	Numero di lotto
Σ	Contains sufficient for <n> tests/</n>	Ausreichend für "n" Ansätze	Contenu suffisant pour "n" tests	Contenido suficiente para <n> ensayos</n>	Contenuto sufficiente per "n" saggi
	Storage Temperature	Lagerungstemperatur	Température de conservation	Temperatura de conservación	Temperatura di conservazione
\square	Expiration Date	Mindesthaltbarkeits- datum	Date limite d'utilisation	Fecha de caducidad	Data di scadenza
***	Legal Manufacturer	Hersteller	Fabricant	Fabricante	Fabbricante
Distributed by	Distributor	Vertreiber	Distributeur	Distribuidor	Distributore
Content	Content	Inhalt	Conditionnement	Contenido	Contenuto
Volume/No.	Volume / No.	Volumen/Anzahl	Volume/Quantité	Volumen/Número	Volume/Quantità