Manufactured for Immuno-Biological Laboratories Inc. (IBL-America) 8201 Central Avenue, NE, Suite P Minneapolis, MN 55432 Tel: 763-780-2955

Toll Free: 1-888-523-1246



Borrelia burgdorferi IgG/IgM ELISA

CONTENTS

- **INTENDED USE** 1
- 2 **BACKGROUND**
- **TEST PRINCIPLE** 3
- 4 **KIT COMPONENTS**
- 5 MATERIAL REQUIRED BUT NOT SUPPLIED
- STORAGE AND STABILITY 6
- **TEST PROCEDURE**
 - 7.1 Evidence of Deterioration
 - 7.2 Sample Preparation and Storage
 - 7.3 Preparation of Kit Reagents
 - 7.4 Overview Test Procedure
 - 7.5 Manual Test Procedure
 - 7.6 Automated Test Procedure
 - 7.7 Positive Control / Accuracy Control
 - 7.8 CSF Testing

TEST EVALUATION

- 8.1 Criteria of Validity
- 8.2 Cut-off Calculation
- 8.3 Borderline Ranges

PERFORMANCE CHARACTERISTICS

- 9.1 Sensitivity and Specificity
- 9.2 Reproducibility

SAFETY MEASURES

- 10.1 Statements of Warning
- 10.2 Disposal

11 **REFERENCES**

For Research Use Only - Not for Use in Clinical **Procedures**

Version No.: V 121.22

IBL-America Borrelia burgdorferi IgG/IgM ELISA

Enzyme-immunoassay for the determination of human antibodies

Borrelia burgdorferi IgG ELISA Order No.: IB05015

Borrelia burgdorferi IgM ELISA Order No.: IB05016

For Research Use Only - Not for Use in Clinical Procedures

1 INTENDED USE

The IBL-America Borrelia burgdorferi IgG and IgM ELISA tests are qualitative immunoassays for the detection of human antibodies in serum, plasma or cerebrospinal fluid directed against *Borrelia burgdorferi sensu lato*.

2 BACKGROUND

Borrelia burgdorferi is the infectious agent which causes the disease syndrome known as Lyme-Borreliosis. B. burgdorferi sensu stricto, B. garinii and B. afzelii are the most important pathogens of the genospecies Borrelia burgdorferi sensu lato. All three are distributed throughout Europe in all temperate climate zones. Reservoirs for the bacteria include a variety of small wild mammals, particularly mice. The bacteria, belonging to the taxonomic group of the spirochetes, are transmitted to hosts by infected ticks (Ixodes ricinus, Europe; Ixodes scapularis, USA). The infection rate in adult ticks, nymphs and larvae with Borrelia burgdorferi ranges up to 50 %, dependent upon geographical area and tick populations. The distribution of the genospecies in infected ticks varies across Europe, however, most ticks are carrying Borrelia afzelli and Borrelia garinii.

3 TEST PRINCIPLE

The ELISA (Enzyme Linked Immunosorbent Assay) is an immunoassay, which is particularly suited to the determination of antibodies in the field of infectious serology. The reaction is based on the specific interaction of antibodies with their corresponding antigen. The test strips of the microtiter plate are coated with specific antigens of the pathogen of interest. If antibodies in the sample are present, they bind to the fixed antigen. A secondary antibody, which has been conjugated with the enzyme alkaline phosphatase, detects and binds to the immune complex. The colourless substrate p-nitrophenylphosphate is then converted into the coloured product p-nitrophenol. The signal intensity of this reaction product is proportional to the concentration of the analyte in the sample and is measured photometrically.

4 KIT COMPONENTS

Test Components	Pieces / Volume
Break apart microtiter test strips each with eight antigen coated single wells, (altogether 96) MTP, 1 frame. The coating material is inactivated.	12 pieces
Standard serum (ready-to-use) STD, Human serum in protein containing phosphate buffer; negative for anti-HIV Ab, HBs-Ag (Hepatitis B-Virus surface antigen) and anti-HCV Ab; preservative: < 0.1 % sodium azide; colouring: Amaranth O	2 x 2 ml
Negative control serum (ready-to-use) NEG, Human serum in protein containing phosphate buffer; negative for anti-HIV Ab, HBs-Ag (Hepatitis B-Virus surface antigen) and anti-HCV Ab; preservative: < 0.1 % sodium azide; colouring: Lissamin Green V	2 ml
Anti-human IgA, IgG or IgM conjugate (ready-to-use) APC, Anti-human IgA, IgG or IgM polyclonal antibody, conjugated to alkaline phosphatase, stabilised with protein stabilisation solution; preservative: < 0.1 % methylisothiazolone, < 0.1 % bromnitrodioxane	13 ml
Washing solution concentrate (sufficient for 1000 ml) WASH, Sodium chloride solution with Tween 20 and 30 mM Tris/HCl, pH 7.4; preservative: < 0.1 % sodium azide	33.3 ml
Special dilution buffer S2 (ready-to-use) DILBS2, Protein containing phosphate buffer with Tween 20 and ultrasonicate of <i>Treponema phagedenis</i> ; preservative: < 0.1 % sodium azide; colouring: 0.01 g/l Bromphenol blue	2 x 50 ml
Stopping solution (ready-to-use) STOP, < 0.1 N sodium hydroxide, 40 mM EDTA	15 ml
Substrate (ready-to-use) pNPP, Para-nitrophenylphosphate in solvent free buffer; preservative: < 0.1 % sodium azide	13 ml
Quality control certificate	1 page

5 MATERIAL REQUIRED BUT NOT SUPPLIED

- Common laboratory equipment
- For the IgM detection: Rf-Absorbent, order no. IB05998 (20 ml)
- Photometer for microtiter plates with filter, wavelength 405 nm, recommended reference wavelength 620 nm 690 nm (e.g. 650 nm)
- Microtiter plate washer
- Incubator 37 °C
- Moist chamber
- Distilled water

6 STORAGE AND STABILITY

Reagent	Storage	Stability
Microtiter strips	unopened	see expiry date
(coated with antigen)	after opening at 2 – 8 °C in closed aluminum bag with desiccant	6 months
Control sera /	unopened	see expiry date
Standard sera	after opening at 2 – 8 °C	6 months
Conjugate	Unopened	see expiry date
	after opening at 2 – 8 °C	6 months
Dilution buffer	Unopened	see expiry date
	after opening at 2 – 8 °C	6 months
Washing solution	unopened / after opening at 2 – 8 °C	see expiry date
	working dilution at 2 – 8 °C	2 weeks
	working dilution at room temperature	1 week
Substrate	Unopened	see expiry date
	after opening at 2 – 8 °C	6 months
Stopping solution	Unopened	see expiry date
	after opening at 2 – 8 °C	6 months

7 TEST PROCEDURE

7.1 Evidence of Deterioration

Optimum results can only be achieved if the instructions are strictly followed. The components must not be exchanged for reagents of other manufacturers. Standard and control sera are defined exclusively for the test kit to be used and must not be used in other lots. Washing solution, substrate and stop solution can be used for all IBL-America immunoassays coded IB05xxx irrespective of the lot and the test.

Each IBL-America ELISA coded IB05xxx contains a ready-to-use sample dilution buffer. In some cases the use of special dilution buffers is necessary to guarantee consistent quality and reliable results. The dilution buffers can be used irrespective of the lots.

There are three different conjugate concentrations for each immunoglobulin class (IgA, IgG, IgM), indicated on the label as + (Iow), ++ (medium) and +++ (high). Conjugates with the same concentration and of the same immunoglobulin class are interchangeable and can be used for other IBL-America ELISAs coded IB05xxx irrespective of the lot and the test. Dilution or alteration of the reagents may result in a loss of sensitivity. Use aseptic techniques when removing aliquots from the reagent tubes to avoid contamination.

Reproducibility of test results is dependent on thorough mixing of the reagents. Agitate the flasks containing control sera before use and also all samples after dilution (e.g. by using a vortex mixer).

Be sure to pipette carefully and comply with the given incubation times and temperatures. Significant time differences between pipetting the first and last well of the microtiter plate when dispensing samples and control sera, conjugate or substrate can result in different pre-incubation times, which may influence the precision and reproducibility of the results. Avoid exposure of reagents to strong light during storage and incubation.

Adequate washing avoids test unspecificities. Therefore, the washing procedure should be carried out carefully. All of the flat bottom wells should be filled with equal volumes of washing buffer. At the end of the procedure ensure that the wells are free of all washing buffer in order to avoid uncontrolled dilution effects. Avoid foaming!

Reagents must be tightly closed after use to avoid evaporation and contamination. Take care not to mix-up the caps of the bottles and/or vials.

The result of this ELISA is only valid if the lot-specific validation criteria on the quality control certificate are fulfilled.

7.2 Sample Preparation and Storage

Lipaemic, hemolytic or icteric samples (serum or plasma) should only be tested with caution. Obviously contaminated samples should not be tested. Serum or plasma (EDTA, citrate, heparin) or CSF collected according to standard laboratory methods are suitable samples. Samples must not be thermally inactivated.

7.2.1 Dilution of Samples

Before running the test, all samples (V_1) must be diluted in dilution buffer (V_2) as follows:

These Borrelia burgdorferi IgG and IgM ELISA testkits include a special dilution buffer B231-S2 containing lysate of *Treponema phagedenis* for the absorption of potentially cross-reacting spirochete-antibodies. Shake bottle with dilution buffer immediately before use!

Borrelia burgdorferi IgG ELISA

$V_1 + V_2 = 1 + 100$	add	10 µl	sample
	each to	1000 µl	dilution buffer

Borrelia burgdorferi IgM ELISA

Interference with rheumatoid factors

Rheumatoid factors are autoantibodies mainly of the IgM class, which preferably bind to IgG immune complexes. The presence of non-specific IgM antibodies (rheumatoid factors) can lead to false-positive results in the IgM assay. Furthermore, the possibility exists, that weak-binding pathogen-specific IgM antibodies may be displaced by stronger-binding IgG antibodies leading to a false-negative IgM result. Therefore it is necessary to pretreat samples with rheumatoid factor-absorbens prior to IgM detection (Rf-Absorbent). Rf-absorption is performed by incubation of the sample in Rf-dilution buffer for 15 minutes at room temperature or over night at 4 °C. The test procedure is described in a separate instruction manual.

Before running the test, rheumatoid factor-absorbent (V_1) must be diluted 1+4 in dilution buffer (V_2) .

$V_1 + V_2 = V_3 (1 + 4)$	add 200 µl	Rf-absorbent
	each to 800 µl	dilution buffer

Samples (V₄) must be diluted in this Rf-dilution buffer (V₃):

$V_4 + V_3 = 1 + 100$	add	10 µl	sample
	each to	1000 µl	Rf-dilution buffer

After dilution and before pipetting into the microtiter plate the samples must be mixed thoroughly to prepare a homogenous solution.

7.2.2 Sample Storage

The samples should not be stored for more than 7 days at 2-8 °C. Extended storage is possible at \leq -20 °C. Avoid repeated freezing and thawing of samples. Diluted samples can be stored at 2-8 °C for one week.

7.3 Preparation of Kit Reagents

Bring all reagents to room temperature before testing.

7.3.1 Microtiter Test Strips

The microtiter test strips labeled with abreviations for pathogen and immunoglobulin class are packed with a desiccant in an aluminum bag. To open the aluminum bag of the microtiter plate please cut off the top of the marked side only, in order to guarantee proper resealing. Take unrequired cavities out of the frame and put them back into the aluminum bag. Close bag carefully to ensure airtight conditions. Do not use the strips if the aluminum bag is damaged or if the bag with remaining strips and desiccant was not properly resealed.

7.3.2 Control Sera / Standard Sera (ready-to-use)

Control and standard sera are ready-to-use and must not be diluted any further. For each test run - independent of the number of microtiter test strips to be used - control and standard sera must be included. Standard and cut off sera should be set up in duplicate. Do not treat control sera with Rf-absorbent.

7.3.3 Anti-human IgA, IgG or IgM AP-Conjugate (ready-to-use)

The required conjugate concentration (+, ++, +++) is indicated on the quality control certificate. Please refer also to the specification on the label. Avoid contamination.

7.3.4 Washing Solution (Concentrate)

Dilute washing buffer concentrate (V_1) 1:30 with aqua dest. to a final volume of V_2 . Bottles used for the working dilution should be cleaned regularly. Discard cloudy solutions. Example:

Buffer concentrate (V ₁)	Final volume (V ₂)	
33.3 ml	1000 ml	
1.0 ml	30 ml	

7.3.5 Dilution Buffer for Samples (ready-to-use)

Discard cloudy solutions.

7.3.6 Substrate (ready-to-use)

Substrate in unopened bottle may have a slightly yellow coloring, which does not reduce the quality of the product! Avoid contamination.

7.3.7 Stopping Solution (ready-to-use)

7.4 Overview - Test Procedure

IBL-America Borrelia burgdorferi IgG/IgM

In case of IgM detection absorption of rheumatoid factor, see No. 7.2.1; Incubation 15 minutes at room temperature or over night at 4°C

> sample dilution¹ 1+100

Pipette diluted samples and ready-to-use control / standard sera into the microtest wells (100 µl)

Ú

INCUBATION 60 min./ 37 °C moist chamber

Û

WASH (4 x 300 μl DIL WASH)²

Pipette conjugate solution $\boxed{\text{APC}}$ (100 μ I)

Û

INCUBATION 30 min./ 37 °C moist chamber

Û

WASH (4 x 300 μ l DIL WASH)²

Pipette substrate solution pNPP (100 μl)

INCUBATION 30 min./ 37 °C moist chamber

Ú

Pipette stopping solution STOP (100 μl)

ΰ

READ EXTINCTION at 405 nm

¹Special dilution buffers for the following IBL-America ELISA tests: Borrelia burgdorferi IgG, IgM, EBV EA IgG

²For manual use:

tap plate at the end of the wash procedure on paper towel.

7.5 Manual Test Procedure

- 1. Place the required number of **cavities in the frame** and prepare a protocol sheet.
- 2. Add each **100 μl of diluted sample or ready-to-use controls** into the appropriate wells of microtiter test strips. Spare one well for substrate blank, e.g.:

Well	Component		
A1	substrate blank		
B1	negative control		
C1	standard serum		
D1	standard serum		
E1	sample 1		
F1	sample 2		

- 3. **Sample incubation** for 60 minutes (+/- 5 min.) at 37 °C (+/- 1°C) in moist chamber
- 4. After incubation **wash** all wells with washing solution (by automated washer or manually):
 - aspirate or shake out the incubation solution
 - fill each well with 300 µl washing solution
 - aspirate or shake out the washing buffer
 - repeat the washing procedure 3 times (altogether 4 times!)
 - dry by tapping the microtiter plate on a paper towel

5. Addition of conjugate

Add 100 µl of the ready-to-use lgG/lgM conjugate to the appropriate wells (except substrate blank)

- 6. **Conjugate incubation** for 30 minutes (+/- 1 min.) at 37 °C (+/- 1 °C) in moist chamber.
- 7. After incubation **wash** all wells with washing solution (see above).
- 8. Addition of substrate

Add 100 μl of ready-to-use substrate solution to each well (including well for substrate blank!)

- 9. **Substrate incubation** for 30 minutes (+/- 1 min.) at 37 °C (+/- 1 °C) in moist chamber.
- 10. Stopping of the reaction

Add 100 µl stopping solution to each well, shake microtiter plate gently to mix.

11. Read extinction

Read optical desity (OD) within 60 minutes at 405 nm against substrate blank, reference wave length between 620 nm and 690 nm (e.g. 650 nm).

7.6 Automated Test Procedure

The ELISA are validated for use with Immunomat (using the following consumables: VT124, VT111, VT112) and suited for processing on similar analyzers. For processing on the Immunomat the current software version including reagent check has to be used. The automated processing is performed analogous to manual use. Please note, that under special working-conditions internal laboratory adaptations of the substrate incubation times may be necessary.

7.7 Positive Control / Accuracy Control

For the periodic verification of the test method, in order to fulfil the requirements of laboratory internal quality management systems, we recommend using IBL-America ELISA controls (cat.-no. IB05xxxCON, see also chapter 5) to determine precision and accuracy of the test runs. The use of these ELISA controls is described in specific instruction manuals.

7.8 CSF Testing

The ELISA testkits for Borrelia burgdorferi IgG and IgM also can be used for the determination of antibodies in CSF samples. The test procedure is described in a separate instruction manual. Please inquire with IBL-America if needed.

8 TEST EVALUATION

For the qualitative interpretation of serum samples a lot specific correction factor as well as a lot specific grey zone is calculated by manufacturer for each kit lot. These values can be found on the lot specific quality certificate included in each test kit.

For test run control a standard serum is used in each individual test run. For this control serum a reference value with a validity range is determined by the quality control of the manufacturer. Within this range a correct cut-off interpretation is ensured.

8.1 Criteria of Validity

The substrate blank must be < 0.25 OD

The negative control must produce a negative test result.

The mean OD-value (after subtraction of the substrate blank!) of the standard serum must be within the validity range, which is given on the lot specific qualitycontrol certificate.

The variation of OD-values of the standard serum may not be higher than 20%.

If these criteria are not met, the test is not valid and must be repeated.

8.2 Cut-off Calculation

A lot specific quality control certificate is included in the test kit so that the obtained OD values can be interpreted qualitatively. The substrate blank must be substracted from all OD values prior to evaluation.

To fix the cut-off ranges multiply the mean value of the measured standard OD with the lot specific correction factor from the quality certificate. Then add and substract the lot specific grey zone percentage mentioned on the quality certificate to obtain the upper and lower cut-off. The following numbers are an example only, the valid data you will find in the lot-specific QC certificate which comes with each kit.

Lot specific correction factor: 0.805. Lot specific grey zone: 15%

If the measured mean absorbance value of the standard serum is 0.84 OD, the range of the cut-off is: Lower cut-off: (0.84 * 0.805) -15% = OD 0.575

Upper cut-off: (0.84 * 0.805) +15% = OD 0.778

8.3 Borderline Ranges

The borderline range indicates the range for borderline test results. Values obtained, when testing a sample, which fall below this range indicate a negative test result; values above the borderline range are interpreted positive. In cases where the results are within the borderline range a definitive interpretation of the result is not possible. In such cases, the test should be repeated in parallel with a follow-up sample taken one to two weeks later (serum pair).

9 PERFORMANCE CHARACTERISTICS

9.1 Sensitivity and Specificity

The IBL-America Borrelia burgdorferi IgG was was verified in an internal study. The sensitivity exceeded 93% and specificity exceeded 99%.

The IBL-America Borrelia burgdorferi IgM was was verified in an internal study. The sensitivity exceeded 94% and specificity exceeded 98%.

9.2 Reproducibility

Borrelia burgdorferi IgG ELISA:

Sample	Mean Value (OD)	Intraassay CV (%)	Mean Value (OD)	Interassay CV (%)
Sample 1	0.413	1.8	0.435	4.4
Sample 2	1.105	1.9	1.165	3.8
Sample 3	1.889	2.5	1.903	3.6

Borrelia burgdorferi IgM ELISA:

Sample	Mean Value (OD)	Intraassay CV (%)	Mean Value (OD)	Interassay CV (%)
Sample 1	0.743	4.8	0.870	4.4
Sample 2	2.156	2.5	2.442	2.7
Sample 3	2.772	1.7	3.000	1.7

9.3 Cross-reactivities

Borrelia burgdorferi IgG:

To determine detection of cross-reactive antibodies directed against different parameters sera were analyzed with Borrelia burgdorferi IgG and a commercially available anti-Borrelia burgdorferi IgG ELISA. Positive sera (10 sera each) for Cytomegalovirus IgG, Epstein-Barr Virus VCA IgG, Influenza A Virus IgG, Influenza B Virus IgG, Leptospira IgG and Treponema pallidum IgG have been tested as well as sera positive for Rheumatoid factor (RF) and anti-nuclear antibodies (ANA). Within this internal evaluation potential cross-reactivities with five Leptospira IgG, four Treponema pallidum IgG and three RF positive serum samples have been observed which have been confirmed by positive results of the reference assay (with exception of one RF sample). Other cross-reactivities cannot be ruled out in general.

Borrelia burgdorferi IgM:

To determine detection of cross-reactive antibodies directed against different parameters sera were analyzed with Borrelia burgdorferi IgM and a commercially available anti-Borrelia burgdorferi IgM ELISA. Positive sera (10 sera each) for Cytomegalovirus IgM, Epstein-Barr Virus VCA IgM, Influenza A Virus IgM, Influenza B Virus IgM, Leptospira IgM and Treponema pallidum IgM have been tested as well as sera positive for Rheumatoid factor (RF) and anti-nuclear antibodies (ANA). Within this internal evaluation potential cross-reactivities with six Cytomegalovirus IgM, two Epstein-Barr Virus VCA IgM, two Leptospira IgM, three Treponema pallidum IgM, two Herpes Simplex Virus 1/2 IgM, one ANA and three RF positive samples have been observed. Most reactivities have been confirmed by positive or borderline results of the reference assay. Other cross-reactivities cannot be ruled out in general.

9.4 Interfering Substances

Borrelia burgdorferi lgG/lgM:

To determine the influence of interfering substances, sera with different reactivites were analyzed with the Borrelia burgdorferi IgG/IgM ELISA. No interferences have been detected for sera with concentrations up to 2.00 g/L hemoglobin, 11.50 g/L lipemia/triglyceride oder 0.201 g/L bilirubin (conjugated and unconjugated).

10 SAFETY MEASURES

10.1 Statements of Warning

The IBL-America ELISA test kits are designed for use by qualified personnel who are familiar with good laboratory practice. All kit reagents and human specimens should be handled carefully, using established good laboratory practice.

- This kit contains human blood components. Although all control- and cut-off sera have been tested and found negative for anti-HIV-ab, HBs-Ag (*Hepatitis B-Virus-surface Antigen*) and anti-HCV-ab, they should be considered potentially infectious.
- Do not pipette by mouth.
- Do not smoke, eat or drink in areas in which specimens or kit reagents are handled.

- Wear disposable gloves, laboratory coat and safety glasses while handling kit reagents or specimens. Wash hands thoroughly afterwards.
- Sample material and other potentially infectious material should be decontaminated after the test run.
- Reagents should be stored safely and be inaccessible to unauthorized access e.g. children.

10.2 Disposal

Please observe the relevant statutory requirements!

11 REFERENCES

- [1] Aguero-Rosenfeld, M.E., Wang, G., Schwartz, I., Wormser, G.P. (2005) Diagnosis of Lyme borreliosis. Clin. Microbiol. Rev. *18*, 484-509.
- [2] Brouqui, P., Bacellar, F., Baranton, G., Birtles, R.J., Bjoërsdorff, A., Blanco, J.R., Caruso, G., Cinco, M., Fournier, P.E., Francavilla, E., Jensenius, M., Kazar, J., Laferl, H., Lakos, A., Lotric Furlan, S., Maurin, M., Oteo, J.A., Parola, P., Perez-Eid, C., Peter, O., Postic, D., Raoult, D., Tellez, A., Tselentis, Y., Wilske, B; ESCMID Study Group on Coxiella, Anaplasma, Rickettsia and Bartonella; European Network for Surveillance of Tick-Borne Diseases (2004) Guidelines for the diagnosis of tick-borne bacterial diseases. Clin. Microbiol. Infect. *10*, 1108-32.
- [3] Fingerle, V., Wilske, B. (2007) Mikrobiologische Diagnostik der Lyme-Borreliose. J. Lab. Med. 31, 141-8.
- [4] Goettner, G., Schulte-Spechtel, U., Hillermann, R., Liegl, G., Wilske, B., Fingerle, V. (2005) Improvement of Lyme borreliosis serodiagnosis by a newly developed recombinant immunoglobulin G (IgG) and IgM line immunoblot assay and addition of VIsE and DbpA homologues. J.Clin. Microbiol. *43*, 3602-9.
- [5] Liang, F.T., Alvarez, A.L., Gu, Y., Nowling, J.M., Ramamoorthy, R., Philipp, M.T., (1999)
 An Immunodominant Concerved Region within the Variable Domain of VIsE, the Variable Surface Antigen of Borrelia burgdorferi. J Immunol. 163, 5566-73.
- [6] Stanek, G. *et al.* (1996) European Union concerted action on risk assessment in Lyme borreliosis: clinical case definitions for Lyme borreliosis. Wien. Klin. Wochenschr. *108*, 741-7.
- [7] Tewald, F., Braun, R. (1998) Durchführung und Interpretation serologischer Tests bei Verdacht auf Borrelieninfektion. Clin. Lab. *44*, 897-902.
- [8] Wilske, B. (2003) Diagnosis of Lyme Borreliosis in Europe. Vector Borne Zoonotic Dis. 3, 215-27.
- [9] Wilske, B. *et al.* (2000) MIQ12 Lyme-Borreliose. Qualitätsstandards in der mikrobiologisch-infektiologischen Diagnostik. Urban & Fischer Verlag, München Jena, 159. *English version*: http://nr2-borrelien.lmu.de.
- [10] Wilske, B., Schrifer, M.: Borrelia. Manual of Clinical Microbiology (8th edition). ASM Press, Washington D.C. (2003), 937-54.

Manufactured for Immuno-Biological Laboratories Inc. (IBL-America)
8201 Central Avenue, NE, Suite P
Minneapolis, MN 55432
Tel: 763-780-2955

Toll Free: 1-888-523-1246

