Instruction for Use Code No. 27751

Human Tenascin-C Large(FNIII-C) Assay Kit – IBL

96 Well

Please read carefully this instruction prior you use this assay kit.

INSTRUCTIONS FOR USE

This product is for research use only and is not intended for diagnostic use.

KIT COMPONENT

1	Precoated plate: (Anti-Human Tenascin-C (1904MS) Mouse IgG A.P.) 96VVell X 1
2	Labeled antibody conc.:	
	(30X) HRP conjugated Anti-Human Tenascin-C (4F10TT) Mouse IgG Fab'A	.P.) 0.4mL x 1
3	Standard: (Purified Human Tenascin-C)	0.5mL x 2
4	EIA buffer	30mL x 1
5	Solution for labeled antibody	12mL x 1
6	Chromogen:	15mL x 1
7	Stop solution	12mL x 1
8	Wash buffer conc.	50mL x 1

MEASURING SAMPLES

Human serum, heparin plasma and cell culture supernatant.

The plasma samples collected by EDTA, Citrate or NaF tend to give rather low data compare to assay by serum or heparin plasma.

PRINCIPLE

This kit is a solid phase sandwich ELISA (Enzyme-linked Immunosorbent Assay). As a primary antibody is coated on a plate, samples and standard are added into the wells for 1st reaction. After the reaction, HRP-conjugated secondary antibody is added into the wells for 2nd reaction. After washing away unbound the secondary antibody, Tetra Methyl Benzidine (TMB) is added to the wells and color develops.

OPERATING PRECATION

- 1 Test samples should be measured soon after collection. For storage of samples, store them frozen and do not repeat freeze/thaw cycles. Thaw the test samples at a low temperature and mix them completely before measurement.
- 2 Test samples should be diluted with "4, EIA buffer" contained in this kit.
- 3 Duplicate measurement of test samples and standards is recommended.
- 4 Standard curve should run for each assay.
- 5 Use test samples in neutral pH range. The contaminations of organic solvent may affect the measurement.
- 6 All reagents should be brought to room temperature (R.T.) and mixed completely and gently before use. After mixing them, make sure of no change in quality of the reagents.
- 7 Use only "8, Wash buffer conc." contained in this kit for washing the precoated plate. Insufficient washing may lead to the failure in measurement.
- 8 Wash the plate immediately after each reaction using by a plate washer with setting wait time zero second. The O.D. value tends to be lower if washing time is getting longer. If you use a multichannel pipette or a washing bottle due to no availability of any plate washer, filling wash buffer in each well and immediately turn the plate upside down and shake it off to completely remove the wash buffer. Repeat the number of times of wash defined in a table for measurement procedure described in section 3. It should be properly washed off as instructed in order to avoid any insufficient wash.
- 9 Carefully tap the plate against a clean paper towel without contacting with inside of each well to completely remove the washing buffer after repeated the determined number of wash.
- 10 "6, Chromogen TMB solution" should be stored in the dark due to its sensitivity against light. It should be also avoided contact with metals. Required quantity should be prepared into a collecting container for each use.
- After adding TMB solution into the wells, the liquid in the wells gradually changes the color in blue. In this process the plate should be in dark. Remained TMB solution in the collecting container should not be returned into the original bottle of TMB solution to avoid contamination.
- 12 Measurement of O.D. should be done within 30 minutes after addition of "7, Stop solution".

OPERATION MANUAL AND DOSAGES

1. Materials needed but not supplied.

Plate reader
Test tubes for dilution
Deionized water
Paper towel
Refrigerator
Incubator (37°C±1°C)

Micropipette and tip
Measuring cylinder and beaker
Plate washer
Collecting container
(i.e. clean disposable test tube)

2. Preparation

(1) Preparation of wash buffer

Dilute "8, Wash buffer conc." 40 fold with deionized water. The diluted one is used for the assay as a wash buffer. Adjust the required quantities if needed.

(2) Preparation of labeled antibody

Dilute "2, Labeled antibody conc." 30 fold with "5, Solution for labeled antibody" using a prepared collecting container.

(3) Preparation of standard

Add 0.5 mL of deionized water into the vial of "3, Standard" and completely dissolve it. Concentration of the standard is 48 ng/mL. The standards enclosed in this kit can be frozen and stored after reconstitution. However the freeze-thaw shall not be repeated.

Prepare 7 test tubes for dilution of the standard and adding 230 μ L of the EIA buffer into each tube.

Put 230 µL of 48 ng/mL standard into the tube 24 ng/mL (Tube-1) and gently mix it. Afterword, put 230 µL of the mixed liquid of tube-1 into the tube 12 ng/mL (Tube-2) and gently mix it. Dilute two fold standard solution in series to set up 7 points of diluted standard between 24 ng/mL and 0.38 ng/mL.

The standards enclosed in this kit can be frozen and stored after reconstitution. However the freeze-thaw shall not be repeated.

Tube-1	24	ng/mL
Tube-2	12	ng/mL
Tube-3	6	ng/mL
Tube-4	3	ng/mL
Tube-5	1.5	ng/mL
Tube-6	0.75	ng/mL
Tube-7	0.38	na/ml

(4) Preparation of test samples

Dilute test samples with "4, EIA buffer" contained in this kit as follows.

Human serum and heparin plasma: more than 32 fold.*

Cell culture supernatant: more than 4 fold.

3 MEASUREMENT PROCEDURE

(1) Add test sample blank

Determine wells for test sample blank. Put 100µL each of "4, EIA buffer" into the wells.

(2) Add prepared test samples and standard

Put 100 μL prepared test samples and 100 μL prepared standard into appropriate wells.

- (3) Incubation with plate lid (1st reaction).
- (4) Washing

Wash the plate with the prepared wash buffer and remove all liquid.

(5) Add prepared labeled antibody

Put 100 µL prepared labeled antibody into the wells.

- (6) Incubation with plate lid (2nd reaction).
- (7) Washing

Wash the plate with the prepared wash buffer and remove all liquid completely.

(8) Add "6, Chromogen - TMB solution"

Put 100 µL the TMB solution into the wells.

- (9) Incubation in dark
- (10) Add "7, Stop solution"

Put 100 μL the Stop solution into the wells.

(11) Determination of optical density (O.D.)

Remove any dirt or drop of water on the bottom of the plate and confirm there is no bubble on the surface of the liquid. Then, measure the both O.D. of standard and the test samples against a test sample blank.

Measurement wavelength: 450 nm. In case of 2 wavelengths:

Main wavelength is 450nm. Sub-wavelength is between 600 and 650 nm.

Table for measurement procedure*

	Test samples	Standard	Test sample blank
Reagents	Test samples 100 μL	Diluted Standard 100 µL	EIA buffer 100 μL
1 st reaction	Incubation for 60 minutes at 37°C with plate lid.		°C with plate lid.
Washing	(Refer	han 350 μL) escribed in TION.)	
Labeled antibody	100 μL	100 µL	100 μL
2 nd reaction	ction Incubation for 30 minutes at		~8°C with plate lid.
Washing	5 times (wash buffer more than 350 μL) (Refer to No. 8 and 9 described in OPERATING PRECATION.)		
TMB solution	100 μL	100 μL	100 µL
Chromogenic reaction	Incubation for 30 minutes at R.T. (shielded).		
Stop solution	100 μL	100 µL	100 µL
Measuring O.D.	O.D. 450 nm / 600~650 nm		nm

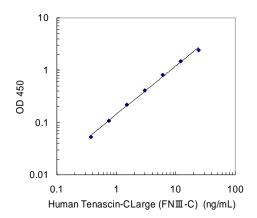


CALCULATION OF TEST RESULT

- 1 Plot the concentration of the standard on the x-axis and its O.D. on the y-axis. Draw a standard curve by applying appropriate regression curve on each plot (i.e. quadratic regression of double logarithm conversion).
- 2 Read the concentration by applying the absorbance of the test samples on a standard curve
- 3 Calculate the concentration of the test samples by multiplying dilution ratio of test samples on the value.

Example of standard curve and measured value

Standard (ng/mL)	O.D. (450nm)
24	2.373
12	1.463
6	0.798
3	0.387
1.5	0.195
0.75	0.086
0.38	0.032



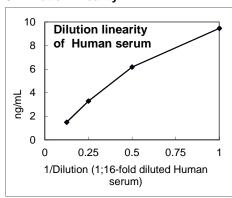
PERFORMANCE AND CHARACTERISTICS

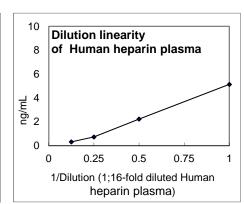
1 Sensitivity

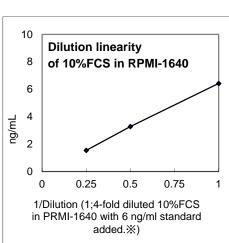
0.01 ng/mL (Calculated by NCCLS method using the standard.)

2 Measurement range 0.38 ∼ 24 ng/mL

3 Dilution linearity*







(XSamples with standard added are used.)

4 Added recovery assay

Specimen	Additive Amount (ng/mL)	Theoretical Value (ng/mL)	Measurement Value (ng/mL)	%		
10%FCS in	12.00	12.03	11.51	95.7		
RPMI-1640	6.00	6.03	5.45	90.4		
(x4)	3.00	3.03	2.86	94.4		
Human	1.50	4.98	5.38	108.0		
Serum	0.75	4.23	4.80	113.5		
(x32)	0.38	3.86	4.06	105.2		
Human	1.50	2.88	3.43	119.1		
heparin	0.75	2.13	2.42	113.6		
Plasma(x32)	0.38	1.76	2.06	117.0		

5 Intra-assay

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Measurement value (ng/ml)	SD(ng/ml)	CV (%)	n	
13.54	0.52	3.8	22	
3.22	0.14	4.3	22	
0.91	0.06	6.6	22	

6 Inter-assay

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Measurement value (ng/ml)	SD (ng/ml)	CV (%)	n
13.55	0.52	3.8	3
3.25	0.17	5.2	3
0.93	0.07	7.5	3

7 Specificity

Specifically detect Tenascin-C in human serum, heparin-plasma and cell culture supernatant.

PRECAUTION FOR INTENDED USE AND/OR HANDLING

1 Precaution for handling (Hazard prevention)

- (1) Treat the components carefully and wash hands after handling it.
- (2) "7, Stop solution" is a strong acid substance (1N Sulfuric acid). Therefore, it should be careful for the treatment and do not contact your skin and clothes with it. It also needs to pay attention to the disposal of it.

2 Precaution for intended use

- (1) "3, Standard" is lyophilized products. It should be careful to open this vial.
- (2) All reagents should be stored at 2 8°C.
- (3) Precipitation can be seen in "4, EIA buffer", "5, Solution for labeled antibody" and "8, Wash buffer conc.", however, it does not affect its performance.
- (4) Do not mix or replace the reagents with the reagents from a different lot or kit
- (5) Do not use expired reagents.

3 Precaution for disposal

(1) Dispose used materials after rinsing them with large quantity of water.

REFERENCE*

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- Tanaka H et al. Circulating level of large splice variants of tenascin-C is a marker of piecemeal necrosis activity in patients with chronic hepatitis C. Liver Int. 2006 Apr; 26 (3):311-8.
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STORAGE AND THE TERM OF VALIDITY

Storage Condition: 2 - 8°C

The expiry date is specified on the outer box.

PACKAGE UNIT AND PRODUCT NUMBER

Package unit: 96 Well Product number: 27751

CONTACT DETAILS*



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