

Mouse/Rat Amyloid β (1-40) High Specific Assay Kit - IBL

96 Well

Please read carefully this instruction prior you use this assay kit.

INSTRUCTIONS FOR USE

This product is for research use only and is not intended for diagnostic use.

KIT COMPONENT

1	Precoated plate: (Anti-A β (35-40)(1A10)Mouse IgG MoAb A.P.)	96Well x 1
2	Labeled antibody conc.: (30X) HRP conjugated Anti-m/rA β (1-16) Rabbit IgG Fab' A.P.)	0.4mL x 1
3	Standard: (Mouse/Rat A β (1-40))	0.5mL x 2
4	EIA buffer	30mL x 1
5	Solution for labeled antibody	12mL x 1
6	Chromogen: TMB solution	15mL x 1
7	Stop solution	12mL x 1
8	Wash buffer conc.	50mL x 1

MESUREING SAMPLES

Mouse/Rat brain extract, plasma, serum and cell culture supernatant.

<Preparation of brain extracted solution>

Add 5 volumes of extraction buffer (1% CHAPS in TBS pH7.6) to brain sample, and homogenize them. After the homogenization, let stand the emulsion on ice for at least 3 hours. Centrifuge it at 70,000 rpm for 20 minutes at 4°C, and then appropriately dilute the supernatant with EIA buffer contained in the kit, and use for measurement. In this case, using the same extraction buffer for reconstruction of standard is recommended.

PRINCIPLE

This kit is a solid phase sandwich ELISA (Enzyme-linked Immunosorbent Assay). As a primary antibody is coated on a plate, samples and standard are added into the wells for 1st reaction. After the reaction, HRP-conjugated secondary antibody is added into the wells for 2nd reaction. After washing away unbound the secondary antibody, Tetra Methyl Benzidine (TMB) is added to the wells and color develops.

OPERATING PRECATION

- Test samples should be measured soon after collection. For storage of samples, store them frozen and do not repeat freeze/thaw cycles. Thaw the test samples at a low temperature and mix them completely before measurement.
- Test samples should be diluted with "4, EIA buffer" contained in this kit.
- Duplicate measurement of test samples and standards is recommended.
- Standard curve should run for each assay.
- Use test samples in neutral pH range. The contaminations of organic solvent may affect the measurement.
- All reagents should be brought to room temperature (R.T.) and mixed completely and gently before use. After mixing them, make sure of no change in quality of the reagents.
- Use only "8, Wash buffer conc." contained in this kit for washing the precoated plate. Insufficient washing may lead to the failure in measurement.
- Fill the wash buffer each well, invert the plate and make sure the liquid is completely removed by shaking it off if you use a washing bottle. Repeat this washing process several times as instructed in order to avoid any insufficient washing process.
- After remove the wash buffer, tapping the plate against a clean paper towel for completely removing the liquid from the wells and make sure the paper towel is not contact with inside of the wells in this process.
- "6, Chromogen - TMB solution" should be stored in the dark due to its sensitivity against light. It should be also avoided contact with metals. Required quantity should be prepared into a collecting container for each use.
- After adding TMB solution into the wells, the liquid in the wells gradually changes the color in blue. In this process the plate should be in dark. Remained TMB solution in the collecting container should not be returned into the original bottle of TMB solution to avoid contamination.
- Measurement of O.D. should be done within 30 minutes after addition of "7, Stop solution".
- When the culture supernatant contains FCS, A β (1-40)-like substances are sometimes measured, and it is recommended that a negative control be set.

OPERATION MANUAL AND DOSAGES

1. Materials needed but not supplied.

Plate reader	Micropipette and tip
Test tubes for dilution	Measuring cylinder and beaker
Deionized water	Plate washer
Paper towel	Collecting container
Refrigerator	(i.e. clean disposable test tube)

2. Preparation

- Preparation of wash buffer
Dilute "8, Wash buffer conc." 40 fold with deionized water. The diluted one is used for the assay as a wash buffer. Adjust the required quantities if needed.
- Preparation of labeled antibody
Dilute "2, Labeled antibody conc." 30 fold with "5, Solution for labeled antibody" using a prepared collecting container.

- Preparation of standard
Add 0.5 mL of deionized water into the vial of "3, Standard" and completely dissolve it. Concentration of the standard is 200 pg/mL. Prepare 7 test tubes for dilution of the standard and adding 230 μ L of the EIA buffer into each tube.

Put 230 μ L of 200 pg/mL standard into the tube 100 pg/mL (Tube-1) and gently mix it. Afterword, put 230 μ L of the mixed liquid of tube-1 into the tube 50 pg/mL (Tube-2) and gently mix it. Dilute two fold standard solution in series to set up 7 points of diluted standard between 100 pg/mL and 1.56 pg/mL.

Tube-1	100	pg/mL	23.6	pmol/L
Tube-2	50	pg/mL	11.8	pmol/L
Tube-3	25	pg/mL	5.9	pmol/L
Tube-4	12.5	pg/mL	2.95	pmol/L
Tube-5	6.25	pg/mL	1.48	pmol/L
Tube-6	3.13	pg/mL	0.74	pmol/L
Tube-7	1.56	pg/mL	0.37	pmol/L
Molecular weight of A β (1-40) is 4233.8				

- Preparation of test samples
Test sample should be diluted with "4, EIA buffer" accordingly.

3 MEASUREMENT PROCEDURE

- Add test sample blank
Determine wells for test sample blank. Put 100 μ L each of "4, EIA buffer" into the wells.
- Add prepared test samples and standard
Put 100 μ L prepared test samples and 100 μ L prepared standard into appropriate wells.
- Incubation with plate lid (1st reaction).
- Washing
Wash the plate with the prepared wash buffer and remove all liquid.
- Add prepared labeled antibody
Put 100 μ L prepared labeled antibody into the wells.
- Incubation with plate lid (2nd reaction).
- Washing
Wash the plate with the prepared wash buffer and remove all liquid completely.
- Add "6, Chromogen - TMB solution"
Put 100 μ L the TMB solution into the wells.
- Incubation in dark
- Add "7, Stop solution"
Put 100 μ L the Stop solution into the wells.
- Determination of optical density (O.D.)
Remove any dirt or drop of water on the bottom of the plate and confirm there is no bubble on the surface of the liquid. Then, measure the both O.D. of standard and the test samples against a test sample blank. Measurement wavelength: 450 nm. In case of 2 wavelengths: Main wavelength is 450nm. Sub-wavelength is between 600 and 650 nm.

Table for measurement procedure

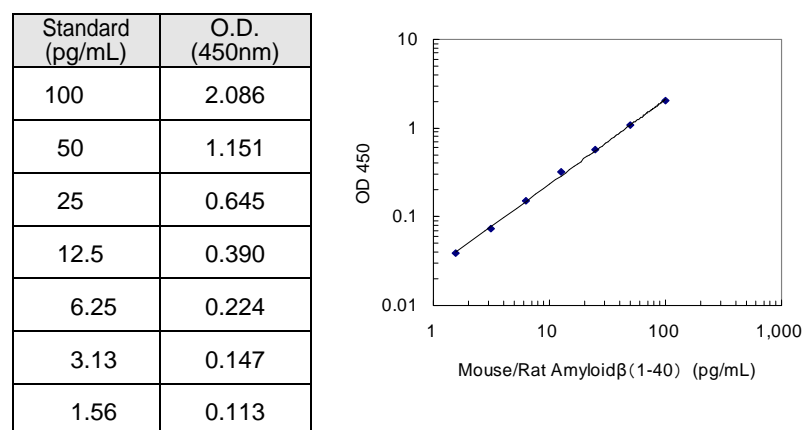
	Test samples	Standard	Test sample blank
Reagents	Test samples 100 μ L	Diluted Standard 100 μ L	EIA buffer 100 μ L
1 st reaction	Incubation for Overnight 2~8°C with plate lid.		
Washing	4 times (wash buffer more than 350 μ L)		
Labeled antibody	100 μ L	100 μ L	100 μ L
2 nd reaction	Incubation for 60 minutes at 2~8°C with plate lid.		
Washing	5 times (wash buffer more than 350 μ L)		
TMB solution	100 μ L	100 μ L	100 μ L
Chromogenic reaction	Incubation for 30 minutes at R.T. (shielded).		
Stop solution	100 μ L	100 μ L	100 μ L
Measuring O.D.	450 nm / 600~650 nm		

CALCULATION OF TEST RESULT

- Plot the concentration of the standard on the x-axis and its O.D. on the y-axis. Draw a standard curve by applying appropriate regression curve on each plot

- (i.e. quadratic regression of double logarithm conversion).
- Read the concentration by applying the absorbance of the test samples on a standard curve.
 - Calculate the concentration of the test samples by multiplying dilution ratio of test samples on the value.

Example of standard curve and measured value



PERFORMANCE AND CHARACTERISTICS

1 Sensitivity

0.28 pg/mL

2 Measurement range

1.56 ~ 100 pg/mL (0.37 ~ 23.6 pmol/L)

3 Dilution test (Samples with standard added are used.)

Test samples	Dilution ratio (x)	Theoretical value (pg/mL)	Measurement value (pg/mL)	%
Plasma (SD rat)	4	13.54	12.16	89.8
	8	7.21	6.36	88.2
	16	3.95	3.41	86.3
Serum (SD rat)	4	13.92	9.50	68.2
	8	6.38	4.47	70.1
	16	2.87	2.34	81.5
Brain Extract (SD rat)	16	14.84	13.84	93.3
	32	8.85	7.99	90.3
	64	4.63	4.34	93.7
Plasma (Balb/c mouse)	4	25.13	21.58	85.9
	8	14.30	11.33	79.2
	16	7.38	5.81	78.7
Serum (Balb/c mouse)	4	17.47	16.75	95.9
	8	7.97	7.20	90.3
	16	3.84	3.35	87.2
Brain Extract (Balb/c mouse)	4	39.56	38.73	97.9
	8	21.57	24.12	111.8
	16	12.34	13.03	105.6

4 Added recovery assay

Test samples	Theoretical value (pg/mL)	Measurement value (pg/mL)	%
Plasma (SD rat) x 4	16.62	11.95	71.9
	10.37	8.21	79.2
	7.24	5.98	82.6
Serum (SD rat) x 4	14.44	7.90	54.7
	11.31	7.59	67.1
	9.75	6.79	69.6
Brain Extract (SD rat) x 8	30.85	28.26	91.6
	27.73	22.56	81.4
	26.16	21.30	81.4
Plasma (Balb/c mouse) x 4	28.65	31.39	109.6
	25.53	24.76	97.0
	23.97	21.61	90.2
Serum (Balb/c mouse) x 4	19.23	15.80	82.2
	16.10	14.40	89.4
	14.54	12.38	85.1
Brain Extract (Balb/c mouse) x8	22.86	22.37	97.9
	16.61	16.15	97.2
	13.49	12.47	92.4

5 Intra-assay

Measurement value (pg/mL)	SD (pg/mL)	CV (%)	n
68.58	2.48	3.6	24
21.10	0.49	2.3	24
4.07	0.20	4.9	24

6 Inter-assay

Measurement value (pg/mL)	SD (pg/mL)	CV (%)	n
69.50	3.02	4.3	24
21.77	1.27	5.8	24
4.04	0.23	5.7	24

7 Specificity

Substance	Cross reactivity (%)
Mouse/Rat A β (1-40)	100
Human A β (1-40)	0.8

PRECAUTION FOR INTENDED USE AND/OR HANDLING

1 Precaution for handling (Hazard prevention)

- Treat the components carefully and wash hands after handling it.
- "7, Stop solution" is a strong acid substance (1N Sulfuric acid). Therefore, it should be careful for the treatment and do not contact your skin and clothes with it. It also needs to pay attention to the disposal of it.

2 Precaution for intended use

- "3, Standard" is lyophilized products. It should be careful to open this vial.
- All reagents should be stored at 2 - 8°C.
- Precipitation can be seen in "4, EIA buffer", "5, Solution for labeled antibody" and "8, Wash buffer conc.", however, it does not affect its performance.
- Do not mix or replace the reagents with the reagents from a different lot or kit.
- Do not use expired reagents.

3 Precaution for disposal

- Dispose used materials after rinsing them with large quantity of water.

STORAGE AND THE TERM OF VALIDITY

Storage Condition: 2 - 8°C

The expiry date is specified on the outer box.

PACKAGE UNIT AND PRODUCT NUMBER

Package unit: 96 Well

Product number: 27720

REFERENCES

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- Saido TC, Iwatsubo T, Mann DM, Shimada H, Ihara Y, Kawashima S. Dominant and differential deposition of distinct beta-amyloid peptide species, A beta N3(pE), in senile plaques. *Neuron.* 1995 Feb;14(2):457-66.

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