Code No. 27407

Human c-Met Assay Kit - MCM

INTRODUCTION

c-Met, the product of *c-met* gene has been reported to be the HGF receptor since HGF induces phosphorylation of c-Met by binding specifically to it. It is a heterodimer protein composed of an α-chain that is linked to a β-chain. The β-chain has the intracellular tyrosine kinase domain, the transmembrane domain and the extracellular domain, and it is tied into the α-chain that is the extracellular domain of c-Met. c-Met exists mainly in epidermal cells. It is found in the digestive tract, prostate gland, seminal vesicles, mammary gland, microglia cells, monocytes and macrophages, and more amount in the liver and kidney. It is considered that c-Met transmits signals resulting from binding to HGF, such as growth, motility and organ formation, in the organs and cells mentioned above.

This kit is designed to measure Human c-Met.

PRINCIPLE

This kit is a solid phase sandwich ELISA using 2 kinds of high specific antibodies. Tetra Methyl Benzidine (TMB) is used as coloring agent (Chromogen). The strength of coloring is in proportion to the quantities of human c-Met.

MEASUREMENT RANGE

0.78 ～ 50 ng/mL

INTENDED USE

For research use only, not for use in diagnostic procedures.

- The IBL's Human c-Met Assay Kit is a complete kit for the quantitative determination of human c-Met in serum, EDTA-plasma or cell culture media.
- Determination of human c-Met is affected by the presence of heparin in samples, so please use EDTA-plasma as a sample instead of heparin plasma.

KIT COMPONENT

1	Precoated plate : Anti- Human c-Met Mouse IgG MoAb Affinity Purify	96Well x 1
2	Labeled antibody Conc. : (30X) HRP conjugated Anti-Human c-Met Mouse IgG Fab' Affinity Purifiy	0.4mL x 1
3	Standard : Human c-Met	0.5mL x 2
4	EIA buffer*	30mL x 1
5	Solution for Labeled antibody*	12mL x 1
6	Chromogen : TMB solution	15mL x 1
7	Stop solution*	12mL x 1
8	Wash buffer Conc.*	50mL x 1

OPERATION MANUAL

1. Materials needed but not supplied

- Plate reader (450nm)
- Micropipette and tip
- Graduated cylinder and beaker
- Deionized water
- Incubator (37℃±1℃)
- Graph paper (log/log)
- Tube for dilution of Standard
- Paper towel
- Refrigerator (as 4℃)
- Washing bottle for precoated plate
- Disposable test tube for "2, Labeled antibody Conc." and "6, Chromogen"

2. Preparation

1) Preparation of wash buffer
"8, Wash buffer Conc." is a concentrated (40X) buffer. The temperature of "8, Wash buffer Conc." shall be adjusted to room temperature and then, mix it gently and completely before use. Dilute 50mL of "8, Wash buffer Conc." with 1,950mL of deionized water and mix it. This is the wash buffer for use. This prepared wash buffer shall be stored in refrigerator and used within 2 weeks after dilution.

2) Preparation of Labeled antibody
"2, Labeled antibody Conc." is a concentrated (30X). Dilute "2, Labeled antibody Conc." with "5, Solution for Labeled antibody" in 30 times according to required quantity into a disposable test tube. Use this resulting solution as Labeled antibody.
Example)
In case you use one slit (8 well), the required quantity of Labeled antibody is 800 μL. (Dilute 30 μL of "2, Labeled antibody Conc." with 870 μL of "5, Solution for Labeled antibody" and mix it. And use the resulting solution by 100 μL in each well.)
This operation should be done just before the application of Labeled antibody.
The remaining "2, Labeled antibody Conc." should be stored at 4℃ in firmly sealed vial.

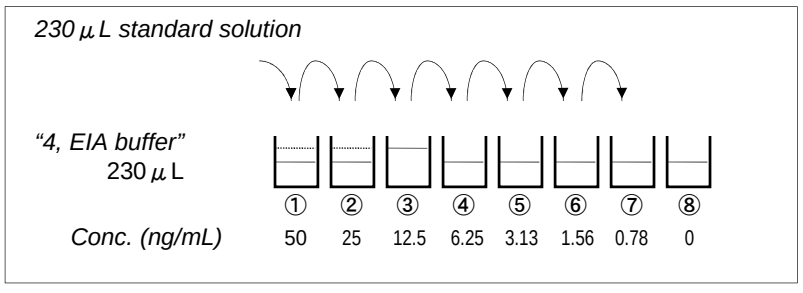
3) Preparation of Standard
Put just 0.5 mL of deionized water into the vial of "3, Standard" and mix it gently and completely. This solution is 100 ng/mL Human c-Met standard.

4) Dilution of Standard
Prepare 8 tubes for dilution of "3, Standard". Put 230 μL each of "4, EIA buffer" into the tube.
Specify the following concentration of each tube.

Tube-1	50 ng/mL
Tube-2	25 ng/mL
Tube-3	12.5 ng/mL
Tube-4	6.25 ng/mL
Tube-5	3.13 ng/mL
Tube-6	1.56 ng/mL
Tube-7	0.78 ng/mL
Tube-8	0 ng/mL (Test Sample Blank)

Put 230 μL of Standard solution into tube–1 and mix it gently. Then, put 230 μL of tube-1 mixture into tube-2. Dilute two times standard solution in series to set up 7 points of diluted standard between 50 ng/mL and 0.78 ng/mL. Tube-8 is the test sample blank as 0 ng/mL.

See following picture.



5) Dilution of test sample
Test sample should be diluted with "4, EIA buffer" accordingly.
If the concentration of Human c-Met in samples may not be estimated in advance, the pre-assay with several different dilutions will be recommended to determine the proper dilution of samples.

3. Measurement procedure

All reagents shall be brought to room temperature approximately 30 minutes before use. Then mix it gently and completely before use. Confirm no change in quality of the reagents. Standard curve shall be prepared simultaneously with the measurement of test samples.

Reagents	Test Sample	Standard	Test Sample Blank	Reagent Blank
	Test sample 100 μL	Diluted standard (Tube 1~7) 100 μL	EIA buffer (Tube-8) 100 μL	EIA buffer 100 μL
Incubation for 60 minutes at 37℃ with plate lid				
4 times (wash buffer more than 350 μL)				
Labeled Antibody	100 μL	100 μL	100 μL	-
Incubation for 30 minutes at 4℃ with plate lid				
5 times (wash buffer more than 350 μL)				
Chromogen	100 μL	100 μL	100 μL	100 μL
Incubation for 30 minutes at room temperature (shielded)				
Stop solution	100 μL	100 μL	100 μL	100 μL
Read the plate at 450nm against a Reagent Blank within 30 minutes after application of Stop solution.				

1) Determine wells for reagent blank. Put 100 μL each of "4, EIA buffer" into the wells.
2) Determine wells for test sample blank, test sample and diluted standard. Then, put 100 μL each of test sample blank (tube-8), test sample and dilutions of standard (tube-1~7) into the appropriate wells.
3) Incubate the precoated plate for 60 minutes at 37℃ after covering it with plate lid.
4) Wash the plate with the prepared wash buffer and remove all liquid.*
5) Pipette 100 μL of labeled antibody solution into the wells of test samples, diluted standard and test sample blank.
6) Incubate the precoated plate for 30 minutes at 4℃ after covering it with plate lid.
7) Wash the plate with the prepared wash buffer and remove all liquid.*
8) "6, Chromogen" should be taken the required quantity into a disposable test tube. Then, pipette 100 μL from the test tube into the wells. Please avoid to return the rest of test tube into "6, Chromogen" bottle due to avoid to cause of contamination.
9) Incubate the precoated plate for 30 minutes at room temperature in the dark. The liquid will turn blue by the addition of "6, Chromogen".
10) Pipette 100 μL of "7, Stop solution" into the wells. Mix the liquid by tapping the side of precoated plate. The liquid will turn yellow by the addition of "7, Stop solution".
11) Remove any dirt or drop of water on the bottom of the precoated plate and confirm there is no bubble on the surface of the liquid. Then, run the plate reader and conduct measurement at 450nm.
The measurement shall be done within 30minutes after the addition of "7, Stop solution".

SPECIAL ATTENTION

1) Test samples should be measured soon after the collection. In case of the storage of test samples, they should be stored under frozen conditions and do not repeat freeze/thaw cycles. Thaw the test samples at low temperature and mix them completely before measurement.
2) Test samples should be diluted with "4, EIA buffer", if the need arises.
3) The measurement of test samples and standard in duplicate is recommended.
4) Use test samples in neutral pH range. The contaminations of organic solvent may affect the measurement.
5) Use only wash buffer contained in this kit for washing the precoated plate. Insufficient washing may lead to the failure in measurement.
6) Remove the wash buffer completely by tapping the precoated plate on paper towel.
Do not wipe wells with paper towel.

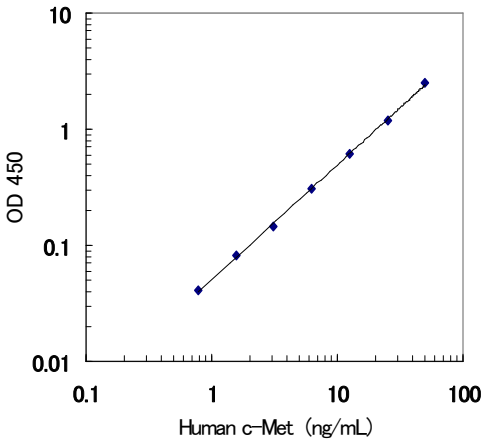
7) “6, Chromogen” should be stored in the dark due to its sensitivity against light. “6, Chromogen” should be avoided contact with metals.
8) Measurement should be done within 30 minutes after addition of “7, Stop solution”.

CALCULATION OF TEST RESULT

Subtract the absorbance of test sample blank from all data, including standards and unknown samples before plotting. Plot the subtracted absorbance of the standards against the standard concentration on log-log graph paper. Draw the best smooth curve through these points to construct the standard curve. Read the concentration for unknown samples from the standard curve.

Example of standard curve

Conc. (ng/mL)	Absorbance (450nm)
50	2.549
25	1.199
12.5	0.617
6.25	0.322
3.13	0.158
1.56	0.092
0.78	0.053
0 (Test Sample Blank)	0.012



* The typical standard curve is shown above. This curve can not be used to derive test results. Please run a standard curve for each assay.

PERFORMANCE CHARACTERISTICS

1. Titer Assay (Samples with standard added are used.)

Specimen	Titer (X)	Measurement Value (ng/mL)	Theoretical Value (ng/mL)	%
10% FBS added RPMI-1640	2	23.03	25.00	92.1
	4	10.93	12.50	87.4
	8	5.74	6.25	91.8
Human Serum	2	19.81	31.82	62.3
	4	11.19	16.74	66.8
	8	6.98	9.07	77.0
Human Plasma (EDTA)	2	23.83	32.50	73.3
	4	12.09	16.81	71.9
	8	7.39	9.00	82.1

2. Added Recovery Assay

Specimen	Theoretical Value (ng/mL)	Measurement Value (ng/mL)	%
10% FCS added RPMI-1640 (x4)	25.09	24.41	97.3
	12.59	10.63	84.4
	6.34	5.14	81.1
Human Serum (x4)	15.37	9.85	64.1
	9.12	6.19	67.9
	5.99	4.60	76.8
Human Plasma (EDTA) (x4)	15.45	9.85	63.8
	9.20	6.56	71.3
	6.08	4.82	79.3

3. Intra - Assay

Measurement Value (ng/mL)	SD value	CV value (%)	n
17.46	0.85	4.9	24
6.09	0.32	5.3	24
2.12	0.13	6.1	24

4. Inter - Assay

Measurement Value (ng/mL)	SD value	CV value (%)	n
18.44	1.09	5.9	36
6.36	0.57	9.0	36
2.26	0.25	11.1	36

5. Specificity

Compound	Cross Reactivity
Human c-Met	100%
Human HGF	≦0.1%
Human HGFA	≦0.1%
Human HAI-1	≦0.1%
Human HAI-2	≦0.1%

6. Sensitivity

0.26 ng/mL
The sensitivity for this kit was determined using the guidelines under the National Committee for Clinical Laboratory Standards (NCCLS) Evaluation Protocols. (National Committee for Clinical Laboratory Standards Evaluation Protocols, SC1, (1989) Villanova, PA: NCCLS.)

PRECAUTION FOR INTENDED USE AND/OR HANDLING

1. All reagents should be stored at 2~8℃. All reagents shall be brought to room temperature approximately 30 minutes before use.
2. “3, Standard” is lyophilized products. Be careful to open this vial.
3. “7, Stop solution” is a strong acid substance. Therefore, be careful not to contact your skin and clothes with “7, Stop solution” and pay attention to the disposal of “7, Stop solution”.
4. Dispose used materials after rinsing them with large quantity of water.
5. The precipitation may grow in “2, Labeled antibody Conc.”, however, there is no problem in the performance.
6. Wash hands after handling reagents.
7. Do not mix the reagents with the reagents from different lot or different kit.
8. Do not use the reagents expired.
9. This kit is for research purpose only. Do not use for clinical diagnosis.

STORAGE AND THE TERM OF VALIDITY

Storage Condition : 2 - 8 °C
The expiry date is specified on outer box.

REFERENCE

1. Kataoka H, Hamasuna R, Itoh H, Kitamura N, Koono M. Activation of hepatocyte growth factor/scatter factor in colorectal carcinoma. Cancer Res. 2000 Nov 1;60(21):6148-59.

2. Shimomura T, Denda K, Kitamura A, Kawaguchi T, Kito M, Kondo J, Kagaya S, Qin L, Takata H, Miyazawa K, Kitamura N. Hepatocyte growth factor activator inhibitor, a novel Kunitz-type serine protease inhibitor. J Biol Chem. 1997 Mar 7;272(10):6370-6.

3. Kawaguchi T, Qin L, Shimomura T, Kondo J, Matsumoto K, Denda K, Kitamura N. Purification and cloning of hepatocyte growth factor activator inhibitor type 2, a Kunitz-type serine protease inhibitor. J Biol Chem. 1997 Oct 31;272(44):27558-64.

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