

Code No. 27361

Human Intelectin-1/Omentin-1 Assay Kit - IBL

INTRODUCTION

Intelectin-1/Omentin-1 is a secretory animal lectin that binds to galactofuranose. Human intelectin-1 is a glycoprotein mainly secreted from intestinal goblet cells. It was reported that the protein was related to inflammation or parasite infection. Recently, it has been reported that intelectin-1 is secreted from epithelioid-type malignant pleural mesothelioma into pleural effusion, while human intelectin-1 is not expressed on normal mesothelial cells or most cancer cells including lung adenocarcinoma (ref. 1). More recently, Intelectin-1/Omentin-1 has been known as an adipocytokine that exists in human blood, and is expected as a new biomarker of metabolic risk factor.

This product is an ELISA kit for measuring of human Intelectin-1/Omentin-1.

PRINCIPLE

This kit is a solid phase sandwich ELISA using 2 kinds of highly specific antibodies. Tetra Methyl Benzidine (TMB) is used as a coloring agent (Chromogen). The strength of coloring is proportional to the quantities of Human Intelectin-1/Omentin-1.

MEASUREMENT RANGE

0.31 - 20 ng/mL

INTENDED USE

For research use only, not for use in diagnostic procedures.

- This assay kit is capable for the quantitative determination human Intelectin-1/Omentin-1 in serum, plasma and cell culture supernatant.
- Heparin-plasma is recommended, EDTA-plasma is not acceptable.
- The guide line of dilution rate for serum and plasma samples is about 100-fold.

KIT COMPONENT

1	Precoated plate Anti-Human Intelectir	: n-1 (2D11) Mouse IgG MoAb Affinity Purify	96Well x 1
2	Labeled antibody Conc.	. ,	
	(30X) HRP conjugated Anti-	Human Intelectin-1 (2D2) Mouse IgG MoAb Fab' Affinity Purify	0.4mL x 1
3	Standard	: recombinant Human Intelectin-1/Omentin-1	0.5mL x 2
4	EIA buffer		50mL x 1
5	Unused number		
6	Chromogen	: TMB solution	15mL x 1
7	Stop solution	: 1N H₂SO₄	12mL x 1
8	Wash buffer Conc.	: (40X) Phosphate buffer	50mL x 1

OPERATION MANUAL

1. Materials needed but not supplied

- Plate reader (450nm)
- · Graduated cylinder and beaker
- Refrigerator (as 4°C)
- · Paper towel
- Incubator (37°C ± 1°C) · Washing bottle for precoated plate
- · Disposable test tube for "2, Labeled antibody Conc." and "6, Chromogen"

Micropipette and tip

Graph paper (log/log)

· Tube for dilution of Standard

· Deionized water

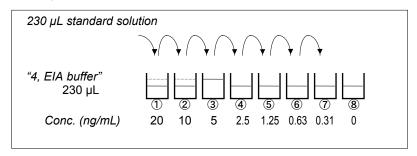
2. Preparation

- 1) Preparation of wash buffer
 - "8, Wash buffer Conc." is a concentrated (40X) buffer. Adjust the temperature of "8, Washing buffer Conc." to room temperature and then, mix it gently and completely before use. Dilute 50 mL of "8, Wash buffer Conc." with 1,950 mL of deionized water and mix it. This is the wash buffer for use. This prepared wash buffer shall be stored in refrigerator and used within 2 weeks after dilution.
- Preparation of Labeled antibody
 - "2, Labeled antibody Conc." is a concentrated (30X). Dilute "2, Labeled antibody Conc." with "4, EIA buffer" in 30 times according to required quantity into a disposable test tube. Use this resulting solution as Labeled antibody. Example)
 - In case you use one strip (8 well), the required quantity of Labeled antibody is 800 μL. (Dilute 30 μL of "2, Labeled antibody Conc." with 870 μL of "4, EIA buffer" and mix it. And use the resulting solution by 100 µL in each well.) This operation should be done just before applying labeled antibody.
 - The remaining "2, Labeled antibody Conc." should be stored at 4°C in firmly sealed vial.
- Preparation of Standard
 - Put just 0.5 mL of deionized water into the vial of "3, Standard" and mix it gently and completely. This solution is 40 ng/mL Human Intelectin-1/Omentin-1 standard.
- Dilution of Standard
 - Prepare 8 tubes for dilution of "3, Standard". Put 230 µL each of "4, EIA buffer" into the tube.
 - Specify the following concentration of each tube."

Tube-1	20 ng/mL	
Tube-2	10 ng/mL	
Tube-3	5 ng/mL	
Tube-4	2.5 ng/mL	
Tube-5	1.25 ng/mL	
Tube-6	0.63 ng/mL	
Tube-7	0.31 ng/mL	
Tube-8	0 ng/mL	(Test Sample Blank)

Put 230 μL of Standard solution into tube-1 and mix it gently. Then, put 230 μL of tube-1 mixture into tube-2. Dilute two times standard solution in series to set up 7 points of diluted standard between 20 ng/mL and 0.31 ng/mL. Tube-8 is the test sample blank as 0 ng/mL.

See following picture.



5) Dilution of test sample

Test samples have to be diluted with "4, EIA buffer".

If the concentration of Human Intelectin-1/Omentin-1 in samples may not be estimated in advance, the pre-assay with several different dilutions will be recommended to determine the proper dilution of samples.

3. Measurement procedure

All reagents shall be brought to room temperature approximately 30 minutes before use. Then mix it gently and completely before use. Make sure of no change in quality of the reagents. Standard curve shall be prepared simultaneously with the measurement of test samples.

	Test Sample	Standard	Test Sample Blank	Reagent Blank	
Reagents	Test sample 100 μL	Diluted standard (Tube 1-7) 100 µL	EIA buffer (Tube-8) 100 μL	EIA buffer 100 μL	
Incubation for 60 minutes at 37°C with plate lid					
		Washing 5 times	;		
Labeled Antibody	100 μL	100 μL	100 μL	-	
Incubation for 30 minutes at 4°C with plate lid					
Washing 5 times					
Chromogen	100 μL	100 μL	100 μL	100 μL	
Incubation for 30 minutes at room temperature (shielded)					
Stop solution	100 μL	100 μL	100 μL	100 μL	
Read the plate at 450nm against a Reagent Blank within 30 minutes after addition of Stop solution.					

- 1) Determine wells for reagent blank. Put 100 µL each of "4, EIA buffer" into the
- Determine wells for test sample blank, test sample and diluted standard. Then, put 100 μL each of test sample blank (tube-8), test sample and dilutions of standard (tube-1-7) into the appropriate wells.
- Incubate the precoated plate for 60 minutes at 37°C after covering it with plate lid.
- Wash each well of the precoated plate 5 times with wash buffer using a washing bottle or a plate washer in following way.
 - After shaking off (or aspiration of) the solution in wells, fill each well with wash buffer and shake off the wash buffer completely from the precoated plate. This procedure must be repeated 5 times. Then, drain the precoated plate completely on paper towel.
 - In case of using a plate washer, we recommend manually washing in the manner mentioned above at least the last one time.
 - Please refer to 5) and 6) in SPECIAL ATTENION below, and be careful not to miss a well.
- Pipette 100 µL of labeled antibody solution into the wells of test samples, diluted standard and test sample blank.
- Incubate the precoated plate for 30 minutes at 4°C after covering it with plate
- Wash the precoated plate 5 times in the same manner as 4).
- Take the required quantity of "6, Chromogen" into a disposable test tube. Then, pipette 100 µL from the test tube into every well. Please do not return the rest of used chromogen in the test tube into "6, Chromogen" bottle in order to avoid contamination.
- Incubate the precoated plate for 30 minutes at room temperature in the dark. The solution of Chromogen will turn blue.
- Add 100 µL of "7, Stop solution" to all wells. Mix the solution by tapping the de of precoated plate. The solution will turn yellow by addit solution".
- 11) Remove any dirt or drop of water on the bottom of the precoated plate and confirm there is no bubble on the surface of the solution. Then, run the plate reader and conduct measurement at 450 nm against a reagent blank. The measurement shall be done within 30 minutes after addition of "7, Stop solution".

SPECIAL ATTENTION

- Test samples should be measured soon after collection. For the storage of test samples, store them frozen and do not repeat freeze/thaw cycles. Thaw the test samples at a low temperature and mix them completely before measurement.
- Test samples have to be diluted with "4, EIA buffer".
- Duplicate measurement of test samples and standard is recommended.
- Use test samples in neutral pH range. The contaminations of organic solvent may affect the measurement.
- Use only wash buffer in this kit for washing the precoated plate.
- Insufficient washing may lead to the failure in measurement.
- Remove the wash buffer completely by tapping the precoated plate on paper

towel. Do not wipe wells with paper towel.

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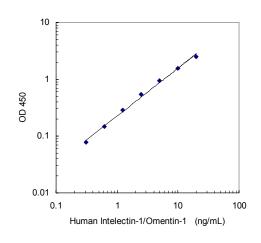
- "6, Chromogen" should be stored in the dark due to its sensitivity against light. Avoid contact of Chromogen with metals.
- Measurement should be done within 30 minutes after addition of "7, Stop solution".

CALCULATION OF TEST RESULT

Subtract the absorbance of test sample blank from all data, including standards and unknown samples before plotting. Plot the subtracted absorbance of the standards against the standard concentration on log-log graph paper. Draw the best smooth curve through these points to construct the standard curve. Read the concentration for unknown samples from the standard curve.

Example of standard curve

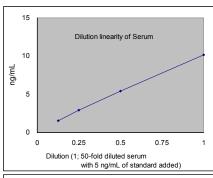
Conc. (ng/mL)	Absorbance (450nm)
20	2.532
10	1.597
5	0.969
2.5	0.565
1.25	0.309
0.63	0.165
0.31	0.096
0 (Test Sample Blank)	0.019

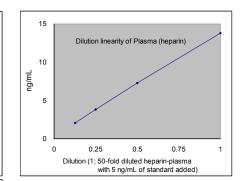


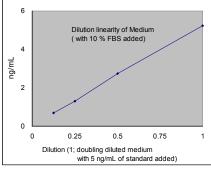
* The typical standard curve is shown above. This curve can not be used to derive test results. Please run a standard curve for each assay.

PERFORMANCE CHARACTERISTICS

1. Dilution linearity







2. Added Recovery Assay

Specimen	Additive Amount (ng/mL)	Theoretical Value (ng/mL)	Measured Value (ng/mL)	%
	5.00	9.15	8.31	90.8
Human Serum	2.50	6.65	6.24	93.8
(x100)	1.25	5.40	4.81	89.1
	0.63	4.77	4.28	89.7
	5.00	10.92	10.06	92.1
Human Plasma (Heparin)	2.50	8.42	7.88	93.6
(x100)	1.25	7.17	6.26	87.3
	0.63	6.54	5.69	87.0
	5.00	5.00	5.50	110.0
Medium with 10% FBS	2.50	2.50	2.33	93.2
(x2)	1.25	1.25	1.19	95.2
	0.63	0.63	0.58	92.1

3. Intra - Assay

Mean Value (ng/mL)	SD (ng/mL)	CV (%)	n
3.14	0.14	4.5	24
1.47	0.08	5.4	24
0.70	0.06	7.8	24

4. Inter - Assay

Mean Value (ng/mL)	SD (ng/mL)	CV (%)	n
3.21	0.23	7.2	7
1.53	0.14	9.2	7
0.68	0.05	7.4	7

5. Sensitivity

0.23 ng/ml

The sensitivity for this kit was determined using the guidelines under the National Committee for Clinical Laboratory Standards (NCCLS) Evaluation Protocols. (National Committee for Clinical Laboratory Standards Evaluation Protocols, SC1, (1989) Villanova, PA: NCCLS.)

PRECAUTION FOR INTENDED USE AND/OR HANDLING

- 1. All reagents should be stored at 2 8°C. All reagents shall be brought to room temperature approximately 30 minutes before use.
- "3, Standard" is lyophilized products. Be careful to open this vial.
- 3. "7, Stop solution" is a strong acid substance. Therefore, be careful not to have your skin and clothes contact "7, Stop solution" and pay attention to the disposal of "7, Stop solution".
- 4. Dispose used materials after rinsing them with large quantity of water.
- 5. Precipitation may occur in "2, Labeled antibody Conc.", "4, EIA buffer" or "8, Wash buffer Conc.", however, there is no problem in the performance.
- 6. Wash hands after handling reagents.
- 7. Do not mix the reagents with the reagents from a different lot or kit.
- 8. Do not use expired reagents.
- 9. This kit is for research purpose only. Do not use for clinical diagnosis.

STORAGE AND THE TERM OF VALIDITY

Storage Condition : 2 - 8°C

The expiry date is specified on outer box.

REFERENCE

- Tsuji S, Tsuura Y, Morohoshi T, Shinohara T, Oshita F, Yamada K, Kameda Y, Ohtsu T, Nakamura Y, Miyagi Y. Secretion of intelectin-1 from malignant pleural mesothelioma into pleural effusion. Br J Cancer. 2010 Aug 10;103(4):517-23.
- 2. Tsuji S, Yamashita M, Hoffman DR, Nishiyama A, Shinohara T, Ohtsu T, Shibata Y. Capture of heat-killed Mycobacterium bovis bacillus Calmette-Guérin by intelectin-1 deposited on cell surfaces. Glycobiology. 2009 May;19(5):518-26.
- 3. Tsuji S, Yamashita M, Nishiyama A, Shinohara T, Li Z, Myrvik QN, Hoffman DR, Henriksen RA, Shibata Y. Differential structure and activity between human and mouse intelectin-1: human intelectin-1 is a disulfide-linked trimer, whereas mouse homologue is a monomer. Glycobiology. 2007 Oct;17(10):1045-51.
- Tsuji S, Uehori J, Matsumoto M, Suzuki Y, Matsuhisa A, Toyoshima K, Seya T. Human intelectin is a novel soluble lectin that recognizes galactofuranose in carbohydrate chains of bacterial cell wall. J Biol Chem. 2001 Jun 29;276(26):23456-63.

Version 1.

Made in Japan.



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