

Code No. 27141

Human SCF Assay Kit - IBL

INTRODUCTION

Stem cell factor (SCF) and its receptor c-Kit play an important role in various biologic phases, such as hematopoiesis, reproduction, and regeneration. Studies in white spotting and steel mice have shown that functional SCF and c-Kit are critical in the survival and development of stem cells involved in hematopoiesis, pigmentation and reproduction. Interaction of SCF with c-Kit rapidly induces receptor dimerization and increases in autophosphorylation activity.

PRINCIPLE

This kit is a solid phase sandwich ELISA using 2 kinds of high specific antibodies. Tetra Methyl Benzidine (TMB) is used as a coloring agent (Chromogen). The strength of coloring is proportional to the quantities of Human SCF.

MEASUREMENT RANGE

50 ~ 3,200 pg/mL

INTENDED USE

- This IBL's assay kit is a capable for the quantitative determination of Human SCF in cell culture medium.
- Both recombinant and native forms Human SCF can be detected with the kit.

KIT COMPONENT

1	Precoated plate : Anti-Human SCF (K089) Rabbit IgG Affinity Purify	96Well x 1
2	Labeled antibody Conc.	
	: (30X) HRP conjugated Anti- Human SCF (K151) Rabbit IgG Fab' Affinity Purify	0.4mL x 1
3	Standard : Recombinant Human SCF	0.5mL x 2
4	EIA buffer *	30mL x 1
5	Solution for Labeled antibody *	12mL x 1
6	Chromogen : TMB solution	15mL x 1
7	Stop solution*	12mL x 1
8	Wash buffer Conc. *	50mL x 1

OPERATION MANUAL

1. Materials needed but not supplied

• Incubator $(37^{\circ}C \pm 1^{\circ}C)$

Plate reader (450nm)

Micropipette and tip

Graduated cylinder and beaker
 Deionized water

Graph paper (log/log)

- Tube for dilution of Standard
- Paper towel
- Washing bottle for precoated plate
 Disposable test tube for "2, Labeled antibody Conc." and "6, Chromogen"

2. Preparation

1) Preparation of wash buffer

"8, Wash buffer Conc." is a concentrated (40X) buffer. Adjust the temperature of "8, Washing buffer Conc." to room temperature and then, mix it gently and completely before use. Dilute 50 mL of "8, Wash buffer Conc." with 1,950 mL of deionized water and mix it. This is the wash buffer for use. This prepared wash buffer shall be stored in refrigerator and used within 2 weeks after dilution.

2) Preparation of Labeled antibody

"2, Labeled antibody Conc." is a concentrated (30X). Dilute "2, Labeled antibody Conc." with "5, Solution for Labeled antibody" in 30 times according to required quantity into a disposable test tube. Use this resulting solution as Labeled antibody.

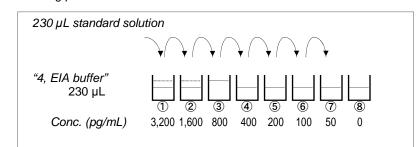
Example)

In case you use one strip (8 well), the required quantity of Labeled antibody is 800 μ L. (Dilute 30 μ L of "2, Labeled antibody Conc." with 870 μ L of "5, Solution for Labeled antibody" and mix it. And use the resulting solution by 100 μ L in each well.)

This operation should be done just before the application of Labeled antibody. The remaining "2, Labeled antibody Conc." should be stored at 4° C in firmly sealed vial.

- 3) Preparation of Standard
- Put just <u>0.5 mL</u> of deionized water into the vial of "3, Standard" and mix it gently and completely. This solution is 6,400 pg/mL Human SCF standard. Dilution of Standard
- Prepare 8 tubes for dilution of "3, Standard". Put 230 µL each of "4, EIA buffer" into the tube.
 - Specify the following concentration of each tube."
 - Tube-1 3 200 ng/ml

See following picture.



5) Dilution of test sample

Test sample should be diluted with "4, EIA buffer" as necessary. If the concentration of Human SCF in samples may not be estimated in advance, the pre-assay with several different dilutions will be recommended to determine the proper dilution of samples.

3. Measurement procedure

All reagents shall be brought to room temperature approximately 30 minutes before use. Then mix it gently and completely before use. Make sure of no change in quality of the reagents. Standard curve shall be prepared simultaneously with the measurement of test samples.

	Test Sample	Standard	Test Sample Blank	Reagent Blank	
Reagents	Test sample 100 µL	Diluted standard (Tube 1~7) 100 μL	EIA buffer (Tube-8) 100 μL	EIA buffer 100 μL	
	Incubation for 60 minutes at 37°C with plate lid				
	4 times (was	h buffer more the	an 350 µL) *		
Labeled Antibody	100 µL	100 µL	100 µL	-	
Incubation for 30 minutes at 37°C with plate lid					
5 times (wash buffer more than 350 µL) *					
Chromogen	100 µL	100 µL	100 µL	100 µL	
Incubation for 30 minutes at room temperature (shielded)					
Stop solution	100 µL	100 µL	100 µL	100 µL	
Read the plate at 450nm against a Reagent Blank within 30 minutes after addition of Stop solution.					

- 1) Determine wells for reagent blank. Put 100 μL each of "4, EIA buffer" into the wells.
- Determine wells for test sample blank, test sample and diluted standard. Then, put 100 µL each of test sample blank (tube-8), test sample and dilutions of standard (tube-1-7) into the appropriate wells.
- 3) Incubate the precoated plate for 60 minutes at 37°C after covering it with plate lid.
- 4) Wash the plate with the prepared wash buffer and remove all liquid. *
- 5) Pipette 100 µL of labeled antibody solution into the wells of test samples, diluted standard and test sample blank.
- 6) Incubate the precoated plate for 30 minutes at 37°C after covering it with plate lid.
- 7) Wash the plate with the prepared wash buffer and remove all liquid. *
- 8) Taken the required quantity of "6, Chromogen" into a disposable test tube. Then, pipette 100µL from the test tube into the wells. Please do not return the rest of the test tube to "6, Chromogen" bottle to avoid contamination.
- Incubate the precoated plate for 30 minutes at room temperature in the dark. The liquid will turn blue by addition of "6, Chromogen".
- 10) Pipette 100 μL of "7, Stop solution" into the wells. Mix the liquid by tapping the side of precoated plate. The liquid will turn yellow by addition of "7, Stop solution".
- 11) Remove any dirt or drop of water on the bottom of the precoated plate and confirm there is no bubble on the surface of the liquid. Then, run the plate reader and conduct measurement at 450 nm against a reagent blank. The measurement shall be done within 30 minutes after addition of "7, Stop solution".

SPECIAL ATTENTION

1) Test samples should be measured soon after collection. For the storage of test samples, store them frozen and do not repeat freeze/thaw cycles. Thaw the

TUDE	5,200 pg/me	
Tube-2	1,600 pg/mL	
Tube-3	800 pg/mL	
Tube-4	400 pg/mL	
Tube-5	200 pg/mL	
Tube-6	100 pg/mL	
Tube-7	50 pg/mL	
Tube-8	0 pg/mL	(Test Sample Blank)

Put 230 μ L of Standard solution into tube-1 and mix it gently. Then, put 230 μ L of tube-1 mixture into tube-2. Dilute two times standard solution in series to set up 7 points of diluted standard between 3,200 pg/mL and 50 pg/mL. Tube-8 is the test sample blank as 0 pg/mL.

test samples at a low temperature and mix them completely before measurement.

- 2) Test samples should be diluted with "4, EIA buffer", if the need arises.
- 3) Duplicate measurement of test samples and standard is recommended.
- 4) Use test samples in neutral pH range. The contaminations of organic solvent may affect the measurement.
- 5) Use only wash buffer contained in this kit for washing the precoated plate. Insufficient washing may lead to the failure in measurement.
- 6) Remove the wash buffer completely by tapping the precoated plate on paper towel. Do not wipe wells with paper towel.
- 7) "6, Chromogen" should be stored in the dark due to its sensitivity against light."6, Chromogen" should be avoided contact with metals.
- 8) Measurement should be done within 30 minutes after addition of "7, Stop solution".

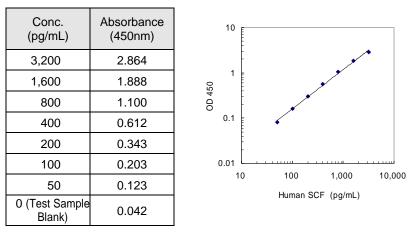
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CALCULATION OF TEST RESULT

Subtract the absorbance of test sample blank from all data, including standards and unknown samples before plotting. Plot the subtracted absorbance of the standards against the standard concentration on log-log graph paper. Draw the best smooth curve through these points to construct the standard curve. Read the concentration for unknown samples from the standard curve.

Example of standard curve



* The typical standard curve is shown above. This curve can not be used to derive test results. Please run a standard curve for each assay.

PERFORMANCE CHARACTERISTICS

1. Titer Assay (Samples with standard added are used.)

Specimen	Titer (X)	Measurement Value (pg/mL)	Theoretical Value (pg/mL)	%
	4	1,621.09	1,604.70	101.0
RPMI-1640 with	8	822.46	803.11	102.4
10 % FCS	16	399.15	401.08	99.5
l	32	189.75	203.43	93.3

2. Added Recovery Assay

Specimen	Theoretical Value (pg/mL)	Measurement Value (pg/mL)	%
RPMI-1640	1,604.68	1,785.77	111.3
with	804.68	925.86	115.1
10 % FCS (x4)	404.68	442.07	109.2

3. Intra - Assay

Measurement Value (pg/mL)	SD value	CV value (%)	n
1,141.20	55.52	4.9	26
262.40	16.77	6.4	26
58.72	3.80	6.5	26

4. Inter - Assay

Measurement Value (pg/mL)	SD value	CV value (%)	n
1,138.63	56.21	4.9	32
258.46	17.64	6.8	32
57.06	5.06	8.9	32

PRECAUTION FOR INTENDED USE AND/OR HANDLING

- 1. All reagents should be stored at 2 8°C. All reagents shall be brought to room temperature approximately 30 minutes before use.
- 2. "3, Standard" is lyophilized products. Be careful to open this vial.
- 3. "7, Stop solution" is a strong acid substance. Therefore, be careful not to have your skin and clothes contact "7, Stop solution" and pay attention to the disposal of "7, Stop solution".
- 4. Dispose used materials after rinsing them with large quantity of water.
- 5. Precipitation may occur in "2, Labeled antibody Conc.", however, there is no problem in the performance.
- 6. Wash hands after handling reagents.
- 7. Do not mix the reagents with the reagents from a different lot or kit.
- 8. Do not use expired reagents.
- 9. This kit is for research purpose only. Do not use for clinical diagnosis.

STORAGE AND THE TERM OF VALIDITY

Storage Condition : 2 - 8°C The expiry date is specified on outer box.

Version 2.

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5. Specificity

Compound	Cross Reactivity
Human SCF	100 %
Human G-CSF	≦ 0.1 %
Human GROα	≦ 0.1 %
Human VEGF (165)	≦0.1 %
Human VEGF (121)	≦0.1 %

6. Sensitivity

7.14 pg/mL

The sensitivity for this kit was determined using the guidelines under the National Committee for Clinical Laboratory Standards (NCCLS) Evaluation Protocols. (National Committee for Clinical Laboratory Standards Evaluation Protocols, SC1, (1989) Villanova, PA: NCCLS.)

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