

**Gd-IgA1 (Galactose-deficient IgA1) Assay Kit-IBL**

96 Well

Please read carefully this instruction prior you use this assay kit.

**INSTRUCTIONS FOR USE**

This product is for research use only and is not intended for diagnostic use.

**KIT COMPONENT**

|   |   |            |
|---|---|------------|
| 1 | <b>Precoated plate:</b> (Anti-Gd-IgA1(KM55) rat IgG.)                                   | 96Well x 1 |
| 2 | <b>Labeled antibody conc.:</b><br>(30X) HRP conjugated Anti-Human IgA(38B1) Rat IgG HRP | 0.2mL x 1  |
| 3 | <b>Standard:</b> (Human IgA1)   | 0.3mL x 2  |
| 4 | <b>EIA buffer</b>   | 30mL x 1   |
| 5 | <b>Solution for labeled antibody</b>  | 6mL x 1    |
| 6 | <b>Chromogen:</b> TMB solution  | 15mL x 1   |
| 7 | <b>Stop solution</b>  | 12mL x 1   |
| 8 | <b>Wash buffer conc.</b>  | 50mL x 1   |

**MEASURING SAMPLES**

Human serum EDTA-plasma and Urine\*

**PRINCIPLE**

This kit is a solid phase sandwich ELISA (Enzyme-linked Immunosorbent Assay). As a primary antibody is coated on a plate, samples and standard are added into the wells for 1<sup>st</sup> reaction. After the reaction, HRP-conjugated secondary antibody is added into the wells for 2<sup>nd</sup> reaction. After washing away unbound the secondary antibody, Tetra Methyl Benzidine (TMB) is added to the wells and color develops.

**OPERATING PRECATION**

- Test samples should be measured soon after collection. For storage of samples, store them frozen and do not repeat freeze/thaw cycles. Thaw the test samples at a low temperature and mix them completely before measurement.
- Test samples should be diluted with "4, EIA buffer" contained in this kit.
- Duplicate measurement of test samples and standards is recommended.
- Standard curve should run for each assay.
- Use test samples in neutral pH range. The contaminations of organic solvent may affect the measurement.
- All reagents should be brought to room temperature (R.T.) and mixed completely and gently before use. After mixing them, make sure of no change in quality of the reagents.
- Use only "8, Wash buffer conc." contained in this kit for washing the precoated plate. Insufficient washing may lead to the failure in measurement.
- Fill the wash buffer each well, invert the plate and make sure the liquid is completely removed by shaking it off if you use a washing bottle. Repeat this washing process several times as instructed in order to avoid any insufficient washing process.
- After remove the wash buffer, tapping the plate against a clean paper towel for completely removing the liquid from the wells and make sure the paper towel is not contact with inside of the wells in this process.
- "6, Chromogen - TMB solution" should be stored in the dark due to its sensitivity against light. It should be also avoided contact with metals. Required quantity should be prepared into a collecting container for each use.
- After adding TMB solution into the wells, the liquid in the wells gradually changes the color in blue. In this process the plate should be in dark. Remained TMB solution in the collecting container should not be returned into the original bottle of TMB solution to avoid contamination.
- Measurement of O.D. should be done within 30 minutes after addition of "7, Stop solution".

**OPERATION MANUAL AND DOSAGES**

**1. Materials needed but not supplied.**

|                         |   |
|-------------------------|---|
| Plate reader            | Micropipette and tip                                      |
| Test tubes for dilution | Measuring cylinder and beaker                             |
| Deionized water         | Plate washer  |
| Paper towel             | Collecting container<br>(i.e. clean disposable test tube) |

**2. Preparation**

- Preparation of wash buffer  
Dilute "8, Wash buffer conc." 40 fold with deionized water. The diluted one is used for the assay as a wash buffer. Adjust the required quantities if needed.
- Preparation of labeled antibody  
Dilute "2, Labeled antibody conc." 30 fold with "5, Solution for labeled antibody" using a prepared collecting container.

(3) Preparation of standard

Add 0.3 mL of deionized water into the vial of "3, Standard" and completely dissolve it. Concentration of the standard is 200 ng/mL. The standards enclosed in this kit can be frozen and stored after reconstitution. However the freeze-thaw shall not be repeated. Prepare 7 test tubes for dilution of the standard and adding 120 µL of the EIA buffer into each tube.

Put 120 µL of 200 ng/mL standard into the tube 100 ng/mL (Tube-1) and gently mix it. Afterword, put 120 µL of the mixed liquid of tube-1 into the tube 50 ng/mL (Tube-2) and gently mix it. Dilute two fold standard solution in series to set up 7 points of diluted standard between 100 ng/mL~1.56 ng/mL.

|        |       |       |
|--------|-------|-------|
| Tube-1 | 100   | ng/mL |
| Tube-2 | 50    | ng/mL |
| Tube-3 | 25    | ng/mL |
| Tube-4 | 12.50 | ng/mL |
| Tube-5 | 6.25  | ng/mL |
| Tube-6 | 3.13  | ng/mL |
| Tube-7 | 1.56  | ng/mL |

(4) Preparation of test samples\*

Dilute test samples with "4, EIA buffer" contained in this kit as follows.

|                   |                 |
|-------------------|-----------------|
| Human serum       | : 200~800 fold. |
| Human EDTA-plasma | : 200~800 fold. |
| Human Urine       | : 2~4 fold*.    |

**3 MEASUREMENT PROCEDURE**

- Add test sample blank  
Determine wells for test sample blank. Put 50 µL each of "4, EIA buffer" into the wells.
- Add prepared test samples and standard  
Put 50 µL prepared test samples and 50 µL prepared standard into appropriate wells.
- Incubation with plate lid (1st reaction).
- Washing  
Wash the plate with the prepared wash buffer and remove all liquid.
- Add prepared labeled antibody  
Put 50 µL prepared labeled antibody into the wells.
- Incubation with plate lid (2<sup>nd</sup> reaction).
- Washing  
Wash the plate with the prepared wash buffer and remove all liquid completely.
- Add "6, Chromogen - TMB solution"  
Put 50 µL the TMB solution into the wells.
- Incubation in dark
- Add "7, Stop solution"  
Put 50 µL the Stop solution into the wells.
- Determination of optical density (O.D.)  
Remove any dirt or drop of water on the bottom of the plate and confirm there is no bubble on the surface of the liquid. Then, measure the both O.D. of standard and the test samples against a test sample blank.  
Measurement wavelength: 450 nm. In case of 2 wavelengths:  
Main wavelength is 450nm. Sub-wavelength is between 600 and 650 nm.

Table for measurement procedure

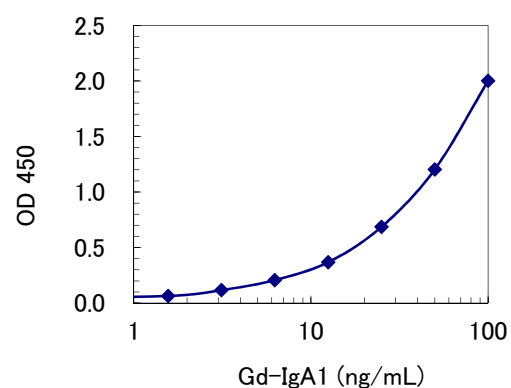
|                          | Test samples                                      | Standard                     | Test sample blank   |
|--------------------------|---|------------------------------|---------------------|
| Reagents                 | Test samples<br>50 µL                             | Diluted<br>Standard<br>50 µL | EIA buffer<br>50 µL |
| 1 <sup>st</sup> reaction | Incubation for 60 minutes at R.T. with plate lid. |                              |                     |
| Washing                  | 4 times (wash buffer more than 350 µL)            |                              |                     |
| Labeled antibody         | 50 µL   | 50 µL                        | 50 µL               |
| 2 <sup>nd</sup> reaction | Incubation for 30 minutes at R.T. with plate lid. |                              |                     |
| Washing                  | 5 times (wash buffer more than 350 µL)            |                              |                     |
| TMB solution             | 50 µL   | 50 µL                        | 50 µL               |
| Chromogenic reaction     | Incubation for 30 minutes at R.T. (shielded).     |                              |                     |
| Stop solution            | 50 µL   | 50 µL                        | 50 µL               |
| Measuring O.D.           | 450 nm / 600~650 nm                               |                              |                     |

**CALCULATION OF TEST RESULT**

- Plot the concentration of the standard on the x-axis and its O.D. on the y-axis. Draw a standard curve by applying appropriate regression curve on each plot (i.e. 4 parameter logistics).
- Read the concentration by applying the absorbance of the test samples on a standard curve.
- Calculate the concentration of the test samples by multiplying dilution ratio of test samples on the value.

Example of standard curve and measured value

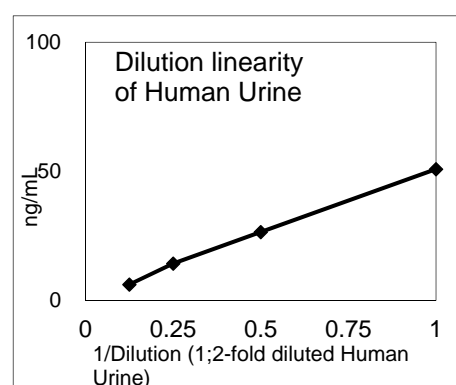
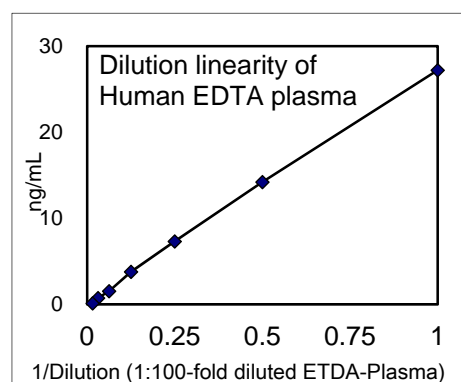
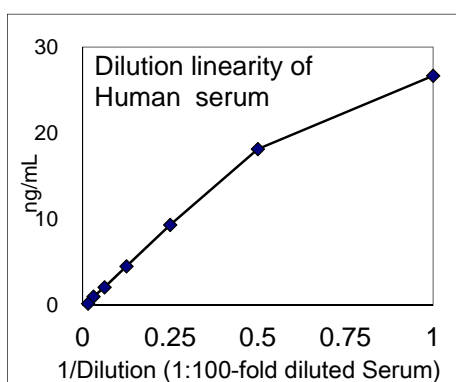
| Standard (ng/mL) | O.D. (450nm) |
|------------------|--------------|
| 100              | 2.001        |
| 50               | 1.204        |
| 25               | 0.686        |
| 12.50            | 0.368        |
| 6.25             | 0.207        |
| 3.13             | 0.117        |
| 1.56             | 0.064        |


**PERFORMANCE AND CHARACTERISTICS**
**1 Sensitivity**

0.488 ng/mL

**2 Measurement range**

1.56 ~ 100 ng/mL

**3 Dilution linearity\***

**4 Added recovery assay\***

| Specimen                 | Additive Amount (ng/mL) | Theoretical Value (ng/mL) | Measurement Value (ng/mL) | %     |
|--------------------------|-------------------------|---------------------------|---------------------------|-------|
| Human Serum (x250)       | 50.00                   | 67.62                     | 60.98                     | 90.2  |
|                          | 25.00                   | 42.62                     | 37.92                     | 89.0  |
|                          | 12.50                   | 30.12                     | 29.14                     | 96.7  |
| Human EDTA-Plasma (x250) | 50.00                   | 67.12                     | 55.73                     | 83.0  |
|                          | 25.00                   | 42.12                     | 35.37                     | 84.0  |
|                          | 12.50                   | 29.62                     | 34.44                     | 116.3 |
| Human Urine (x2)         | 50.00                   | 52.43                     | 45.52                     | 86.8  |
|                          | 25.00                   | 27.43                     | 24.31                     | 88.6  |
|                          | 12.50                   | 14.93                     | 13.59                     | 91.0  |

**5 Intra-assay**

| Measurement value (ng/mL) | SD (ng/mL) | CV (%) | n  |
|---------------------------|------------|--------|----|
| 53.09                     | 4.60       | 8.7    | 21 |
| 22.95                     | 2.11       | 9.2    | 21 |
| 14.55                     | 1.44       | 9.9    | 21 |

**6 Inter-assay**

| Measurement value (ng/mL) | SD (ng/mL) | CV (%) | n |
|---------------------------|------------|--------|---|
| 62.68                     | 7.45       | 11.9   | 5 |
| 26.18                     | 2.38       | 9.1    | 5 |
| 16.25                     | 1.22       | 7.5    | 5 |

**PRECAUTION FOR INTENDED USE AND/OR HANDLING**
**1 Precaution for handling (Hazard prevention)**

- Treat the components carefully and wash hands after handling it.
- "7, Stop solution" is a strong acid substance (1N Sulfuric acid). Therefore, it should be careful for the treatment and do not contact your skin and clothes with it. It also needs to pay attention to the disposal of it.

**2 Precaution for intended use**

- "3, Standard" is lyophilized products. It should be careful to open this vial.
- All reagents should be stored at 2 - 8°C.
- Precipitation can be seen in "4, EIA buffer", "5, Solution for labeled antibody" and "8, Wash buffer conc.", however, it does not affect its performance.
- Do not mix or replace the reagents with the reagents from a different lot or kit.
- Do not use expired reagents.

**3 Precaution for disposal**

- Dispose used materials after rinsing them with large quantity of water.

**STORAGE AND THE TERM OF VALIDITY**

Storage Condition: 2 - 8°C

The expiry date is specified on the outer box.

**PACKAGE UNIT AND PRODUCT NUMBER**

Package unit: 96 Well

Product number: 27600

**REFERENCES**

- Novel lectin-independent approach to detect galactose-deficient IgA1 in IgA nephropathy. Yasutake J et al. Nephrol Dial Transplant. 2015 Aug;30(8):1315-21.

**CONTACT DETAILS\***


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